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5 **PLATELET DERIVED GROWTH FACTOR (PDGF)**
 NUCLEIC ACID LIGAND COMPLEXES

10 **RELATED APPLICATIONS**

15 This application is a Divisional of U.S. Patent Application Ser. No. 09/851,486, filed
May 8, 2001, now U.S. Patent No. 6,592,918, which is a Divisional of U.S. Patent Application
Serial No. 08/991,743, filed December 16, 1997 which is a Continuation-in-Part of United
States Patent Application Serial No. 08/618,693, filed March 20, 1996, now U.S. Patent No.
20 5,723,594, which is a Continuation-in-Part of United States Patent Application Serial No.
08/479,783, filed June 7, 1995, now U.S. Patent No. 6,028,186, and 08/479,725, filed June 7,
1995, now U.S. Patent No. 5,674,685.

25 **FIELD OF THE INVENTION**

20 Described herein are high affinity ssDNA and RNA ligands to platelet derived growth
factor (PDGF). The method utilized herein for identifying such Nucleic Acid Ligands is called
SELEX, an acronym for Systematic Evolution of Ligands by Exponential enrichment. Further
included in this invention is a method for preparing a therapeutic or diagnostic Complex
25 comprised of a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight
Compound or a Lipophilic Compound by identifying a PDGF Nucleic Acid Ligand by SELEX
methodology and covalently linking the PDGF Nucleic Acid Ligand with a Non-Immunogenic,
High Molecular Weight Compound or a Lipophilic Compound. The invention further includes
Complexes comprised of one or more PDGF Nucleic Acid Ligands and a Non-Immunogenic,
30 High Molecular Weight Compound or a Lipophilic Compound. The invention further relates to
improving the Pharmacokinetic Properties of a PDGF Nucleic Acid Ligand by covalently
linking the PDGF Nucleic Acid Ligand with a Non-Immunogenic, High Molecular Weight
Compound or Lipophilic Compound to form a Complex. The invention further relates to
improving the Pharmacokinetic Properties of a PDGF Nucleic Acid Ligand by using a Lipid

Construct comprising a PDGF Nucleic Acid Ligand or a Complex comprising a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound. This invention further relates to a method for targeting a therapeutic or diagnostic agent to a biological target that is expressing PDGF by associating the agent with a Complex comprised of a PDGF Nucleic Acid Ligand and a Lipophilic Compound or Non-Immunogenic, High Molecular Weight Compound, wherein the Complex is further associated with a Lipid Construct and the PDGF Nucleic Acid Ligand is further associated with the exterior of the Lipid Construct.

BACKGROUND OF THE INVENTION

A. The SELEX Process

The dogma for many years was that nucleic acids had primarily an informational role. Through a method known as Systematic Evolution of Ligands by Exponential enrichment, termed SELEX, it has become clear that nucleic acids have three dimensional structural diversity not unlike proteins. SELEX is a method for the *in vitro* evolution of nucleic acid molecules with highly specific binding to target molecules and is described in United States Patent Application Serial No. 07/536,428, filed June 11, 1990, entitled "Systematic Evolution of Ligands by Exponential Enrichment," now abandoned, United States Patent Application Serial No. 07/714,131, filed June 10, 1991, entitled "Nucleic Acid Ligands," now United States Patent No. 5,475,096, and United States Patent Application Serial No. 07/931,473, filed August 17, 1992, entitled "Methods for Identifying Nucleic Acid Ligands," now United States Patent No. 5,270,163 (see also WO 91/19813), each of which is specifically incorporated by reference herein. Each of these applications, collectively referred to herein as the SELEX Patent Applications, describes a fundamentally novel method for making a Nucleic Acid Ligand to any desired target molecule. The SELEX process provides a class of products which are referred to as Nucleic Acid Ligands, each ligand having a unique sequence, and which has the property of binding specifically to a desired target compound or molecule. Each SELEX-identified Nucleic Acid Ligand is a specific ligand of a given target compound or molecule. SELEX is based on the unique insight that Nucleic Acids have sufficient capacity for forming a variety of two- and three-dimensional structures and sufficient chemical versatility available within their monomers

to act as ligands (form specific binding pairs) with virtually any chemical compound, whether monomeric or polymeric. Molecules of any size or composition can serve as targets.

The SELEX method involves selection from a mixture of candidate oligonucleotides and step-wise iterations of binding, partitioning and amplification, using the same general selection scheme, to achieve virtually any desired criterion of binding affinity and selectivity. Starting from a mixture of Nucleic Acids, preferably comprising a segment of randomized sequence, the SELEX method includes steps of contacting the mixture with the target under conditions favorable for binding, partitioning unbound Nucleic Acids from those Nucleic Acids which have bound specifically to target molecules, dissociating the Nucleic Acid-target complexes, amplifying the Nucleic Acids dissociated from the Nucleic Acid-target complexes to yield a ligand-enriched mixture of Nucleic Acids, then reiterating the steps of binding, partitioning, dissociating and amplifying through as many cycles as desired to yield highly specific high affinity Nucleic Acid Ligands to the target molecule.

It has been recognized by the present inventors that the SELEX method demonstrates that Nucleic Acids as chemical compounds can form a wide array of shapes, sizes and configurations, and are capable of a far broader repertoire of binding and other functions than those displayed by Nucleic Acids in biological systems.

The present inventors have recognized that SELEX or SELEX-like processes could be used to identify Nucleic Acids which can facilitate any chosen reaction in a manner similar to that in which Nucleic Acid Ligands can be identified for any given target. In theory, within a Candidate Mixture of approximately 10^{13} to 10^{18} Nucleic Acids, the present inventors postulate that at least one Nucleic Acid exists with the appropriate shape to facilitate each of a broad variety of physical and chemical interactions.

The basic SELEX method has been modified to achieve a number of specific objectives. For example, United States Patent Application Serial No. 07/960,093, filed October 14, 1992, entitled "Method for Selecting Nucleic Acids on the Basis of Structure," describes the use of SELEX in conjunction with gel electrophoresis to select Nucleic Acid molecules with specific structural characteristics, such as bent DNA. United States Patent Application Serial No. 08/123,935, filed September 17, 1993, entitled "Photoselection of Nucleic Acid Ligands," describes a SELEX based method for selecting Nucleic Acid Ligands containing photoreactive

groups capable of binding and/or photocrosslinking to and/or photoinactivating a target molecule. United States Patent Application Serial No. 08/134,028, filed October 7, 1993, entitled "High-Affinity Nucleic Acid Ligands That Discriminate Between Theophylline and Caffeine," now United States Patent No. 5,580,737, describes a method for identifying highly specific Nucleic Acid Ligands able to discriminate between closely related molecules, which can be non-peptidic, termed Counter-SELEX. United States Patent Application Serial No. 08/143,564, filed October 25, 1993, entitled "Systematic Evolution of Ligands by Exponential Enrichment: Solution SELEX," now United States Patent No. 5,567,588, describes a SELEX-based method which achieves highly efficient partitioning between oligonucleotides having high and low affinity for a target molecule.

The SELEX method encompasses the identification of high-affinity Nucleic Acid Ligands containing modified nucleotides conferring improved characteristics on the ligand, such as improved *in vivo* stability or improved delivery characteristics. Examples of such modifications include chemical substitutions at the ribose and/or phosphate and/or base positions. SELEX-identified Nucleic Acid Ligands containing modified nucleotides are described in United States Patent Application Serial No. 08/117,991, filed September 8, 1993, entitled "High Affinity Nucleic Acid Ligands Containing Modified Nucleotides," now United States Patent No. 5,660,985, that describes oligonucleotides containing nucleotide derivatives chemically modified at the 5- and 2'-positions of pyrimidines. United States Patent Application Serial No. 08/134,028, *supra*, describes highly specific Nucleic Acid Ligands containing one or more nucleotides modified with 2'-amino (2'-NH₂), 2'-fluoro (2'-F), and/or 2'-O-methyl (2'-OMe). United States Patent Application Serial No. 08/264,029, filed June 22, 1994, entitled "Novel Method of Preparation of Known and Novel 2' Modified Nucleosides by Intramolecular Nucleophilic Displacement," describes oligonucleotides containing various 2'-modified pyrimidines.

The SELEX method encompasses combining selected oligonucleotides with other selected oligonucleotides and non-oligonucleotide functional units as described in United States Patent Application Serial No. 08/284,063, filed August 2, 1994, entitled "Systematic Evolution of Ligands by Exponential Enrichment: Chimeric SELEX," now United States Patent No. 5,637,459, and United States Patent Application Serial No. 08/234,997, filed April 28, 1994,

entitled "Systematic Evolution of Ligands by Exponential Enrichment: Blended SELEX," now U.S. Patent No. 5,683,867, respectively. These applications allow the combination of the broad array of shapes and other properties, and the efficient amplification and replication properties, of oligonucleotides with the desirable properties of other molecules.

5 The SELEX method further encompasses combining selected nucleic acid ligands with lipophilic compounds or non-immunogenic, high molecular weight compounds in a diagnostic or therapeutic complex as described in United States Patent Application Serial No. 08/434,465, filed May 4, 1995, entitled "Nucleic Acid Ligand Complexes," now U.S. Patent No. 6,011,020. VEGF Nucleic Acid Ligands that are associated with a Lipophilic Compound, such as diacyl
10 glycerol or dialkyl glycerol, in a diagnostic or therapeutic complex are described in United States Patent Application Serial No. 08/739,109, filed October 25, 1996, entitled "Vascular Endothelial Growth Factor (VEGF) Nucleic Acid Ligand Complexes," now U.S. Patent No. 5,859,228. VEGF Nucleic Acid Ligands that are associated with a Lipophilic Compound, such as a glycerol lipid, or a Non-Immunogenic, High Molecular Weight Compound, such as
15 polyethylene glycol, are further described in United States Patent Application Serial No. 08/897,351, filed July 21, 1997, entitled "Vascular Endothelial Growth Factor (VEGF) Nucleic Acid Ligand Complexes," now U.S. Patent No. 6,051,698. VEGF Nucleic Acid Ligands that are associated with a non-immunogenic, high molecular weight compound or lipophilic compound are also further described in PCT/US97/18944, filed October 17, 1997, entitled
20 "Vascular Endothelial Growth Factor (VEGF) Nucleic Acid Ligand Complexes." Each of the above described patent applications which describe modifications of the basic SELEX procedure are specifically incorporated by reference herein in their entirety.

B. LIPID CONSTRUCTS

25 Lipid Bilayer Vesicles are closed, fluid-filled microscopic spheres which are formed principally from individual molecules having polar (hydrophilic) and non-polar (lipophilic) portions. The hydrophilic portions may comprise phosphato, glycerylphosphato, carboxy, sulfato, amino, hydroxy, choline or other polar groups. Examples of lipophilic groups are saturated or unsaturated hydrocarbons such as alkyl, alkenyl or other lipid groups. Sterols
30 (e.g., cholesterol) and other pharmaceutically acceptable adjuvants (including anti-oxidants

like alpha-tocopherol) may also be included to improve vesicle stability or confer other desirable characteristics.

Liposomes are a subset of these bilayer vesicles and are comprised principally of phospholipid molecules that contain two hydrophobic tails consisting of fatty acid chains.

5 Upon exposure to water, these molecules spontaneously align to form spherical, bilayer membranes with the lipophilic ends of the molecules in each layer associated in the center of the membrane and the opposing polar ends forming the respective inner and outer surface of the bilayer membrane(s). Thus, each side of the membrane presents a hydrophilic surface while the interior of the membrane comprises a lipophilic medium. These membranes may
10 be arranged in a series of concentric, spherical membranes separated by thin strata of water, in a manner not dissimilar to the layers of an onion, around an internal aqueous space. These multilamellar vesicles (MLV) can be converted into small or Unilamellar Vesicles (UV), with the application of a shearing force.

The therapeutic use of liposomes includes the delivery of drugs which are normally
15 toxic in the free form. In the liposomal form, the toxic drug is occluded, and may be directed away from the tissues sensitive to the drug and targeted to selected areas. Liposomes can also be used therapeutically to release drugs over a prolonged period of time, reducing the frequency of administration. In addition, liposomes can provide a method for forming aqueous dispersions of hydrophobic or amphiphilic drugs, which are normally unsuitable for
20 intravenous delivery.

In order for many drugs and imaging agents to have therapeutic or diagnostic potential, it is necessary for them to be delivered to the proper location in the body, and the liposome can thus be readily injected and form the basis for sustained release and drug delivery to specific cell types, or parts of the body. Several techniques can be employed to
25 use liposomes to target encapsulated drugs to selected host tissues, and away from sensitive tissues. These techniques include manipulating the size of the liposomes, their net surface charge, and their route of administration. MLVs, primarily because they are relatively large, are usually rapidly taken up by the reticuloendothelial system (principally the liver and spleen). UVs, on the other hand, have been found to exhibit increased circulation times,
30 decreased clearance rates and greater biodistribution relative to MLVs.

Passive delivery of liposomes involves the use of various routes of administration, e.g., intravenous, subcutaneous, intramuscular and topical. Each route produces differences in localization of the liposomes. Two common methods used to direct liposomes actively to selected target areas involve attachment of either antibodies or specific receptor ligands to the surface of the liposomes. Antibodies are known to have a high specificity for their corresponding antigen and have been attached to the surface of liposomes, but the results have been less than successful in many instances. Some efforts, however, have been successful in targeting liposomes to tumors without the use of antibodies, see, for example, U.S. Patent No. 5,019,369, U.S. Patent No. 5,441,745, or U.S. Patent No. 5,435,989.

An area of development aggressively pursued by researchers is the delivery of agents not only to a specific cell type but into the cell's cytoplasm and, further yet, into the nucleus. This is particularly important for the delivery of biological agents such as DNA, RNA, ribozymes and proteins. A promising therapeutic pursuit in this area involves the use of antisense DNA and RNA oligonucleotides for the treatment of disease. However, one major problem encountered in the effective application of antisense technology is that oligonucleotides in their phosphodiester form are quickly degraded in body fluids and by intracellular and extracellular enzymes, such as endonucleases and exonucleases, before the target cell is reached. Intravenous administration also results in rapid clearance from the bloodstream by the kidney, and uptake is insufficient to produce an effective intracellular drug concentration. Liposome encapsulation protects the oligonucleotides from the degradative enzymes, increases the circulation half-life and increases uptake efficiency as a result of phagocytosis of the Liposomes. In this way, oligonucleotides are able to reach their desired target and to be delivered to cells *in vivo*.

A few instances have been reported where researchers have attached antisense oligonucleotides to Lipophilic Compounds or Non-Immunogenic, High Molecular Weight Compounds. Antisense oligonucleotides, however, are only effective as intracellular agents. Antisense oligodeoxyribonucleotides targeted to the epidermal growth factor (EGF) receptor have been encapsulated into Liposomes linked to folate via a polyethylene glycol spacer (folate-PEG-Liposomes) and delivered into cultured KB cells via folate receptor-mediated endocytosis (Wang *et al.* (1995) Proc. Natl. Acad. Sci. USA 92:3318-3322). In addition,

aklylene diols have been attached to oligonucleotides (Weiss *et al.*, U.S. Patent No. 5,245,022). Furthermore, a Lipophilic Compound covalently attached to an antisense oligonucleotide has been demonstrated in the literature (EP 462 145 B1).

Loading of biological agents into liposomes can be accomplished by inclusion in the lipid formulation or loading into preformed liposomes. Passive anchoring of oligopeptide and oligosaccharide ligands to the external surface of liposomes has been described (Zalipsky *et al.* (1997) *Bioconjug. Chem.* 8:111:118).

C. PDGF

Platelet-derived growth factor (PDGF) was originally isolated from platelet lysates and identified as the major growth-promoting activity present in serum but not in plasma. Two homologous PDGF isoforms have been identified, PDGF A and B, which are encoded by separate genes (on chromosomes 7 and 22). The most abundant species from platelets is the AB heterodimer, although all three possible dimers (AA, AB and BB) occur naturally. Following translation, PDGF dimers are processed into ~30 kDa secreted proteins. Two cell surface proteins that bind PDGF with high affinity have been identified, α and β (Heldin *et al.* (1981) *Proc. Natl. Acad. Sci.*, 78: 3664; Williams *et al.* (1981) *Proc. Natl. Acad. Sci.*, 79: 5867). Both species contain five immunoglobulin-like extracellular domains, a single transmembrane domain and an intracellular tyrosine kinase domain separated by a kinase insert domain. The functional high affinity receptor is a dimer and engagement of the extracellular domain of the receptor by PDGF results in cross-phosphorylation (one receptor tyrosine kinase phosphorylates the other in the dimer) of several tyrosine residues. Receptor phosphorylation leads to a cascade of events that results in the transduction of the mitogenic or chemotactic signal to the nucleus. For example, in the intracellular domain of the PDGF β receptor, nine tyrosine residues have been identified that when phosphorylated interact with different src-homology 2 (SH2) domain-containing proteins including phospholipase C-g, phosphatidylinositol 3'-kinase, GTPase-activating protein and several adapter molecules like Shc, Grb2 and Nck (Heldin (1995) *Cell* 80: 213). In the last several years, the specificities of the three PDGF isoforms for the three receptor dimers ($\alpha\alpha$, $\alpha\beta$, and $\beta\beta$) has been elucidated. The α -receptor homodimer binds all three PDGF isoforms with high affinity, the

β -receptor homodimer binds only PDGF BB with high affinity and PDGF AB with approximately 10-fold lower affinity, and the $\alpha\beta$ -receptor heterodimer binds PDGF BB and PDGF AB with high affinity (Westermarck & Heldin (1993) *Acta Oncologica* 32:101). The specificity pattern results from the ability of the A-chain to bind only to the α -receptor and of the B-chain to bind to both a and β -receptor subunits with high affinity.

The role of PDGF in proliferative diseases, such as cancer, restenosis, fibrosis, angiogenesis, and wound healing has been established.

PDGF in Cancer

The earliest indication that PDGF expression is linked to malignant transformation came with the finding that the amino acid sequence of PDGF-B chain is virtually identical to that of p28^{sis}, the transforming protein of the simian sarcoma virus (SSV) (Waterfield *et al.* (1983) *Nature* 304:35; Johnsson *et al.* (1984) *EMBO J.* 3:921). The transforming potential of the PDGF-B chain gene and, to a lesser extent, the PDGF-A gene was demonstrated soon thereafter (Clarke *et al.* (1984) *Nature* 308:464; Gazit *et al.* (1984) *Cell* 39:89; Beckmann *et al.* *Science* 241:1346; Bywater *et al.* (1988) *Mol. Cell. Biol.* 8:2753). Many tumor cell lines have since been shown to produce and secrete PDGF, some of which also express PDGF receptors (Raines *et al.* (1990) Peptide Growth Factors and Their Receptors, Springer-Verlag, Part I, p 173). Paracrine and, in some cell lines, autocrine growth stimulation by PDGF is therefore possible. For example, analysis of biopsies from human gliomas has revealed the existence of two autocrine loops: PDGF-B/ β -receptor in tumor-associated endothelial cells and PDGF-A/ α -receptor in tumor cells (Hermansson *et al.* (1988) *Proc. Natl. Acad. Sci.* 85:7748; Hermansson *et al.* (1992) *Cancer Res.* 52:3213). The progression to high grade glioma was accompanied by the increase in expression of PDGF-B and the β -receptor in tumor-associated endothelial cells and PDGF-A in glioma cells. PDGF overexpression may thus promote tumor growth either by directly stimulating tumor cells or by stimulating tumor-associated stromal cells (e.g., endothelial cells). The proliferation of endothelial cells is a hallmark of angiogenesis. Increased expression of PDGF and/or PDGF receptors has also been observed in other malignancies including fibrosarcoma (Smits *et al.* (1992) *Am. J. Pathol.* 140:639) and thyroid carcinoma (Heldin *et al.* (1991) *Endocrinology* 129:2187).

PDGF in Cardiovascular Disease

Percutaneous transluminal coronary angioplasty (PTCA) has become the most common treatment for occlusive coronary artery disease (CAD) involving one or two coronary arteries. In the United States alone about 500,000 procedures are being done annually, with projections of over 700,000 procedures by the year 2000 and about double those amounts worldwide. PTCA, while it involves manipulations inside of coronary arteries, is not considered to be a cardiac surgical intervention. During the most common PTCA procedure, a balloon catheter is threaded through a femoral artery and is positioned within the plaque-laden segment of an occluded coronary vessel; once in place, the balloon is expanded at high pressure, compressing the plaque and increasing the vessel lumen. Unfortunately, in 30-50% of PTCA procedures, reocclusion gradually develops over a period of several weeks or months due to cellular events in the affected vessel wall. Once reocclusion achieves 50% or greater reduction of the original vessel lumen, clinical restenosis is established in the vessel.

In view of the increasing popularity of coronary angioplasty as a less invasive alternative to bypass surgery, restenosis is a serious medical problem. Smooth muscle cells (SMCs) represent a major component of the restenosis lesions. In uninjured arteries, SMCs reside primarily in the medial vessel layer (tunica media). Upon balloon injury that removes the endothelial cells from the intimal layer (tunica intima), SMCs proliferate and migrate into the intima, forming neointimal thickening characteristic of restenosis lesions. When restenosis occurs subsequent to angioplasty, it is usually treated by repeat angioplasty, with or without placement of a stent, or by vascular graft surgery (bypass).

A stent is a rigid cylindrical mesh that, once placed and expanded within a diseased vessel segment, mechanically retains the expanded vessel wall. The stent is deployed by catheter and, having been positioned at the desired site, is expanded in situ by inflation of a high pressure balloon. Being rigid and non-compressible, the expanded stent achieves and maintains a vessel lumen diameter comparable to that of adjacent non-diseased vessel; being pressed tightly into the overlying intima/media, it is resistant to migration within the vessel in response to blood flow. PTCA with stent placement has been compared with PTCA alone

and shown to reduce restenosis to about half and to significantly improve other clinical outcomes such as myocardial infarction (MI) and need for bypass surgery.

There is now considerable evidence that PDGF B-chain is a major contributor to the formation of neointimal lesions. In a rat model of restenosis, the neointimal thickening was inhibited with anti-PDGF-B antibodies (Ferns (1991) *Science* 253:1129-1132; Rutherford *et al.* (1997) *Atherosclerosis* 130:45-51). Conversely, the exogenous administration of PDGF-BB promotes SMC migration and causes an increase in neointimal thickening (Jawien *et al.* (1992) *J. Clin. Invest.* 89:507-511). The effect of PDGF-B on SMCs is mediated through PDGF α -receptor which is expressed at high levels in these cells after balloon injury (Lindner and Reidy (1995) *Circulation Res.* 76:951-957). Furthermore, the degree of neointimal thickening following balloon injury was found to be inversely related to the level of expression of PDGF α -receptor at the site of injury (Sirois *et al.* (1997) *Circulation* 95:669-676).

United States Patent No. 5,171,217 discloses a method and composition for delivery of a drug to an affected intramural site for sustained release in conjunction with or following balloon catheter procedures, such as angioplasty. The drug may be selected from a variety of drugs known to inhibit smooth muscle cell proliferation, including growth factor receptor antagonists for PDGF.

United States Patent No. 5,593,974 discloses methods for treating vascular disorders, such as vascular restenosis, with antisense oligonucleotides. The method is based on localized application of the antisense oligonucleotides to a specific site in vivo. The oligonucleotides can be applied directly to the target tissue in a mixture with an implant or gel, or by direct injection or infusion.

United States Patent No. 5,562,922 discloses a method for preparing a system suitable for localized delivery of biologically active compounds to a subject. The method relates to treating polyurethane coated substrate with a coating expansion solution under conditions that will allow penetration of the biologically active compound throughout the polyurethane coating. Substrates suitable for this invention include, inter alia, metallic stents. Biologically active compounds suitable for use in this invention include, inter alia, lipid-modified oligonucleotides.

Rutherford *et al.* (1997, *Atherosclerosis* 130:45-51) report substantial inhibition of neointimal response to balloon injury in rat carotid artery using a combination of antibodies to PDGF-BB and basic fibroblast growth factor (bFGF).

PDGF in Renal Disease

5 A large variety of progressive renal diseases are characterized by glomerular mesangial cell proliferation and matrix accumulation (Slomowitz *et al.* (1988) *New Eng. J. Med.* 319:1547-1548) which leads to fibrosis. PDGF B-chain appears to have a central role in driving both of these processes given that 1) mesangial cells produce PDGF *in vitro* and various growth factors induce mesangial proliferation via induction of auto- or paracrine
10 PDGF B-chain synthesis; 2) PDGF B-chain and its receptor are overexpressed in many glomerular diseases; 3) infusion of PDGF-BB or glomerular transfection with a PDGF B-chain cDNA can induce selective mesangial cell proliferation and matrix accumulation *in vivo*; and 4) PDGF B-chain or α -receptor knock-out mice fail to develop a mesangium (reviewed in Floege and Johnson (1995) *Miner. Electrolyte Metab.* 21:271-282). In addition
15 to contributing to kidney fibrosis, PDGF is also believed to play a role in fibrosis development in other organs such as lungs and bone marrow and may have other possible disease associations (Raines *et al.* (1990) Experimental Pharmacology, Peptide Growth Factors and Their Receptors, Sporn & Roberts, eds., pp. 173-262, Springer, Heidelberg).

 One study has examined the effect of inhibition of PDGF B-chain in renal disease:
20 Johnson *et al.*, using a neutralizing polyclonal antibody to PDGF, were able to reduce mesangial cell proliferation and matrix accumulation in experimental mesangioproliferative glomerulonephritis (Johnson *et al.* (1992) *J. Exp. Med.* 175:1413-1416). In this model, injection of an anti-mesangial cell antibody (anti-Thy 1.1) into rats resulted in complement-dependent lysis of the mesangial cells, followed by an overshooting reparative phase that
25 resembled human mesangioproliferative nephritis (Floege *et al.* (1993) *Kidney Int. Suppl.* 39:S47-54). Limitations of the study of Johnson *et al.* (Johnson *et al.* (1992) *J. Exp. Med.* 175:1413-1416) included the necessity to administer large amounts of heterologous IgG and a limitation of the study duration to 4 days due to concerns that the heterologous IgG might elicit an immune reaction.

Inhibition of PDGF

Specific inhibition of growth factors, such as PDGF, has become a major goal in experimental and clinical medicine. However, this approach is usually hampered by the lack of specific pharmacological antagonists. Available alternative approaches are also limited, since neutralizing antibodies often show a low efficacy *in vivo* and are usually immunogenic, and given that *in vivo* gene therapy for these purposes is still in its infancy. Currently, antibodies to PDGF (Johnsson *et al.* (1985) Proc. Natl. Acad. Sci. U. S. A. 82:1721-1725; Ferns *et al.* (1991) Science 253:1129-1132; Herren *et al.* (1993) Biochimica et Biophysica Acta 1173:294-302; Rutherford *et al.* (1997) Atherosclerosis 130:45-51) and the soluble PDGF receptors (Herren *et al.* (1993) Biochimica et Biophysica Acta 1173:294-302; Duan *et al.* (1991) J. Biol. Chem. 266:413-418; Teisman *et al.* (1993) J. Biol. Chem. 268:9621-9628) are the most potent and specific antagonists of PDGF. Neutralizing antibodies to PDGF have been shown to revert the SSV-transformed phenotype (Johnsson *et al.* (1985) Proc. Natl. Acad. Sci. U. S. A. 82:1721-1725) and to inhibit the development of neointimal lesions following arterial injury (Ferns *et al.* (1991) Science 253:1129-1132). Other inhibitors of PDGF such as suramin (Williams *et al.* (1984) J. Biol. Chem. 259:287-5294; Betsholtz *et al.* (1984) Cell 39:447-457), neomycin (Vassbotn *et al.* (1992) J. Biol. Chem. 267:15635-15641) and peptides derived from the PDGF amino acid sequence (Engström *et al.* (1992) J. Biol. Chem. 267:16581-16587) have been reported, however, they are either too toxic or lack sufficient specificity or potency to be good drug candidates. Other types of antagonists of possible clinical utility are molecules that selectively inhibit the PDGF receptor tyrosine kinase (Buchdunger *et al.* (1995) Proc. Natl. Acad. Sci. U. S. A. 92:2558-2562; Kovalenko *et al.* (1994) Cancer Res. 54:6106-6114).

SUMMARY OF THE INVENTION

The present invention includes methods of identifying and producing nucleic acid ligands to platelet-derived growth factor (PDGF) and homologous proteins and the nucleic acid ligands so identified and produced. For the purposes of this application, PDGF refers to PDGF AA, AB, and BB isoforms and homologous proteins. Specifically included in the definition are human PDGF AA, AB, and BB isoforms.

Described herein are high affinity ssDNA and RNA ligands to platelet derived growth factor (PDGF). The method utilized herein for identifying such nucleic acid ligands is called SELEX, an acronym for Systematic Evolution of Ligands by Exponential enrichment. Included herein are the evolved ligands that are shown in Tables 2-3, 6-7, and 9 and Figures 1-2, 8A, 8B and 9A. Further included in this invention is a method for preparing a Complex comprised of a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound by the method comprising identifying a Nucleic Acid Ligand from a Candidate Mixture of Nucleic Acids where the Nucleic Acid is a ligand of PDGF by the method of (a) contacting the Candidate Mixture of Nucleic Acids with PDGF, (b) partitioning between members of said Candidate Mixture on the basis of affinity to PDGF, and c) amplifying the selected molecules to yield a mixture of Nucleic Acids enriched for Nucleic Acid sequences with a relatively higher affinity for binding to PDGF, and covalently linking said identified PDGF Nucleic Acid Ligand with a Non-Immunogenic, High Molecular Weight Compound or a Lipophilic Compound. The invention further comprises a Complex comprised of a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or a Lipophilic Compound.

The invention further includes a Lipid Construct comprising a PDGF Nucleic Acid Ligand or a Complex. The present invention further relates to a method for preparing a Lipid Construct comprising a Complex wherein the Complex is comprised of a PDGF Nucleic Acid Ligand and a Lipophilic Compound.

In another embodiment, this invention provides a method for improving the pharmacokinetic properties of a PDGF Nucleic Acid Ligand by covalently linking the PDGF Nucleic Acid Ligand with a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound to form a Complex and administering the Complex to a patient. The invention further relates to a method for improving the pharmacokinetic properties of a PDGF Nucleic Acid Ligand by further associating the Complex with a Lipid Construct.

It is an object of the present invention to provide Complexes comprising one or more PDGF Nucleic Acid Ligands in association with one or more Non-Immunogenic, High Molecular Weight Compounds or Lipophilic Compounds and methods for producing the same. It is a further object of the present invention to provide Lipid Constructs comprising a Complex.

It is a further object of the invention to provide one or more PDGF Nucleic Acid Ligands in association with one or more Non-Immunogenic, High Molecular Weight Compounds or Lipophilic Compounds with improved Pharmacokinetic Properties.

In embodiments of the invention directed to Complexes comprised of a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound, it is preferred that the Non-Immunogenic, High Molecular Weight Compound is Polyalkylene Glycol, more preferably, polyethylene glycol (PEG). More preferably, the PEG has a molecular weight of about 10-80K. Most preferably, the PEG has a molecular weight of about 20-45K. In embodiments of the invention directed to Complexes comprised of a PDGF Nucleic Acid Ligand and a Lipophilic Compound, it is preferred that the Lipophilic Compound is a glycerolipid. In the preferred embodiments of the invention, the Lipid Construct is preferably a Lipid Bilayer Vesicle and most preferably a Liposome. In the preferred embodiment, the PDGF Nucleic Acid Ligand is identified according to the SELEX method.

In embodiments of the invention directed to Complexes comprising a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound covalently linked to a PDGF Nucleic Acid Ligand or Ligands, the PDGF Nucleic Acid Ligand or Ligands can serve in a targeting capacity.

Additionally, the PDGF Nucleic Acid Ligand can be associated through Covalent or Non-Covalent Interactions with a Lipid Construct without being part of a Complex.

Furthermore, in embodiments of the invention directed to Lipid Constructs comprising a PDGF Nucleic Acid Ligand or a Non-Immunogenic, High Molecular Weight or Lipophilic Compound/PDGF Nucleic Acid Ligand Complex where the Lipid Construct is of a type that has a membrane defining an interior compartment such as a Lipid Bilayer Vesicle, the PDGF Nucleic Acid Ligand or Complex in association with the Lipid Construct may be associated with the membrane of the Lipid Construct or encapsulated within the compartment. In embodiments where the PDGF Nucleic Acid Ligand is in association with the membrane, the PDGF Nucleic Acid Ligand can associate with the interior-facing or exterior-facing part of the membrane, such that the PDGF Nucleic Acid Ligand is projecting into or out of the vesicle. In certain embodiments, a PDGF Nucleic Acid Ligand Complex can be passively loaded onto the outside of a preformed Lipid Construct. In embodiments where the Nucleic Acid Ligand is

projecting out of the Lipid Construct, the PDGF Nucleic Acid Ligand can serve in a targeting capacity.

In embodiments where the PDGF Nucleic Acid Ligand of the Lipid Construct serves in a targeting capacity, the Lipid Construct can have associated with it additional therapeutic or diagnostic agents. In one embodiment, the therapeutic or diagnostic agent is associated with the exterior of the Lipid Construct. In other embodiments, the therapeutic or diagnostic agent is encapsulated in the Lipid Construct or associated with the interior of the Lipid Construct. In yet a further embodiment, the therapeutic or diagnostic agent is associated with the Complex. In one embodiment, the therapeutic agent is a drug. In an alternative embodiment, the therapeutic or diagnostic agent is one or more additional Nucleic Acid Ligands.

It is a further object of the present invention to provide a method for inhibiting PDGF-mediated diseases. PDGF-mediated diseases include, but are not limited to, cancer, angiogenesis, restenosis, and fibrosis. Thus, it is a further object of the present invention to provide a method for inhibiting angiogenesis by the administration of a PDGF Nucleic Acid Ligand or a Complex comprising a PDGF Nucleic Acid Ligand and Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound or a Lipid Construct comprising the Complex of the present invention. It is yet a further object of the present invention to provide a method for inhibiting the growth of tumors by the administration of a PDGF Nucleic Acid Ligand or Complex comprising a PDGF Nucleic Acid Ligand and Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound or a Lipid Construct comprising a Complex of the present invention. It is yet a further object of the invention to provide a method for inhibiting fibrosis by the administration of a PDGF Nucleic Acid Ligand or Complex comprising a PDGF Nucleic Acid Ligand and Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound or a Lipid Construct comprising a Complex of the present invention. It is yet a further object of the invention to provide a method for inhibiting restenosis by the administration of a PDGF Nucleic Acid Ligand or Complex comprising a PDGF Nucleic Acid Ligand and Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound or a Lipid Construct comprising a Complex of the present invention.

It is a further object of the invention to provide a method for targeting a therapeutic or diagnostic agent to a biological target that is expressing PDGF by associating the agent with a

Complex comprised of a PDGF Nucleic Acid Ligand and a Lipophilic Compound or Non-Immunogenic, High Molecular Weight Compound, wherein the Complex is further associated with a Lipid Construct and the PDGF Nucleic Acid Ligand is further associated with the exterior of the Lipid Construct.

5 These and other objects, as well as the nature, scope and utilization of this invention, will become readily apparent to those skilled in the art from the following description and the appended claims.

BRIEF DESCRIPTION OF THE FIGURES

10 Figure 1 shows the consensus secondary structure for the sequence set shown in Table 3. R = A or G, Y = C or T, K = G or T, N and N' indicate any base pair.

 Figure 2 shows the minimal ligands 20t, 36t and 41t folded according to the consensus secondary structure motif. [3'T] represents a 3'-3' linked thymidine nucleotide added to reduce 3'-exonuclease degradation.

15 Figures 3A-3C show the binding of minimal high affinity DNA ligands to PDGF AA, PDGF AB, and PDGF BB, respectively. The fraction of ^{32}P 5' end-labeled DNA ligands bound to varying concentrations of PDGF was determined by the nitrocellulose filter binding method. Minimal ligands tested were 20t (o), 36t (?), and 41t (?). Oligonucleotide concentrations in these experiments were ~10 pM (PDGF-AB and PDGF-BB) and ~50 pM (PDGF AA). Data points were fitted to eq. 1 (for binding of the DNA ligands to PDGF-AA) or to eq. 2 (for binding to PDGF AB and BB) using the non-linear least squares method. Binding reactions were done at 37°C in binding buffer (PBSM with 0.01% HSA).

20 Figure 4 shows the dissociation rate determination for the high affinity interaction between the minimal DNA ligands and PDGF AB. The fraction of 5' ^{32}P end-labeled ligands 20t (o), 36t (?), and 41t (?), all at 0.17 nM, bound to PDGF AB (1 nM) was measured by nitrocellulose filter binding at the indicated time points following the addition of a 500-fold excess of the unlabeled competitor. The dissociation rate constant (k_{off}) values were determined by fitting the data points to eq 3 in Example 1. The experiments were performed at 37°C in binding buffer.

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Figure 5 shows the thermal denaturation profiles for the minimal high affinity DNA ligands to PDGF-AB. The change in absorbance at 260 nm was measured in PBS containing 1 mM MgCl₂ as a function of temperature for ligands 20t (o), 36t (?), and 41t (?).

Figure 6 shows the effect of DNA ligands on the binding of ¹²⁵I-PDGF-BB to PDGF
5 a-receptors expressed in PAE cells.

Figure 7 shows the effect of DNA ligands on the mitogenic effect of PDGF-BB on PAE cells expressing the PDGF β-receptors.

Figures 8A-8B show the substitution pattern compatible with high affinity binding to PDGF-AB. In Figures 8A-8C, the underlined symbols indicate
10 2'-O-methyl-2'-deoxynucleotides; italicized symbols indicate 2'-fluoro-2'-deoxynucleotides; normal font indicates 2'-deoxyribonucleotides; [3'T] indicates inverted orientation (3'3') thymidine nucleotide (Glen Research, Sterling, VA); PEG in the loops of helices II and III of Figure 8B indicates pentaethylene glycol spacer phosphoramidite (Glen Research, Sterling, VA) (See Figure 9 for molecular description). Figure 8C shows the predicted secondary
15 structure of a scrambled Nucleic Acid Ligand sequence that was used as a control in Examples 8 and 9. The scrambled region is boxed to accent the overall similarity of the scrambled Nucleic Acid Ligand to the Nucleic Acid Ligand shown in Figure 8B.

Figures 9A-9E show the molecular descriptions NX31975 40K PEG (Figure 9A), NX31976 40K(Figure 9B), hexaethylene glycol phosphoramidite (Figure 9C), pentyl amino
20 linker (Figure 9D), and 40K PEG NHS ester (Figure 9E). The 5' phosphate group shown in the PEG Spacer of Figures 9A and 9B are from the hexaethylene glycol phosphoramidite.

Figure 10 shows the stabilities of DNA (36ta) and modified DNA (NX21568) Nucleic Acid Ligands in rat serum over time at 37°C were compared. 36ta is shown by the symbol √; and NX21568 is shown by the symbol ⊗.

Figure 11 shows that NX31975-40K PEG significantly inhibited (p < 0.05) about
25 50% of the neointima formation in rats based on the intima/media ratio for the control (PBS) and NX31975-40K PEG groups.

Figure 12 shows the effects of NX31975 40K PEG on mitogen-stimulated proliferation of mesangial cells in culture (all mitogens were added at 100 ng/ml final
30 concentration). Scrambled Nucleic Acid Ligand NX31976 and 40K PEG were also tested.

Data are optical densities measured in the XTT assay and are expressed as percentages of baseline, i.e., cells stimulated with medium plus 200 $\mu\text{g/ml}$ 40K PEG (i.e., the amount equivalent to the PEG attached to 50 $\mu\text{g/ml}$ Nucleic Acid Ligand). Results are means \pm SD of 5 separate experiments ($n = 3$ in the case of medium plus 40K PEG; statistical evaluation was therefore confined to NX31975 and scrambled Nucleic Acid Ligand groups).

Figures 13A-13E show effects of NX31975 40K PEG on glomerular cell proliferation (Figure 13A), expression of glomerular PDGF B-chain (Figure 13B), proteinuria in rats with anti-Thy 1.1 nephritis (Figure 13C), mesangial cell activation (as assessed by glomerular *de novo* expression of α -smooth muscle actin) (Figure 13D), and monocyte/macrophage influx (Figure 13E). NX31975 40K PEG is shown as black, NX31976 40K PEG is shown as cross-hatched, 40K PEG is shown as white, PBS is shown as hatched, and the normal range is shown as stippled.

Figure 14A-C show the effects of NX31975 40K PEG on glomerular matrix accumulation. Glomerular immunostaining scores for fibronectin and type IV collagen as well as glomerular scores for type IV collagen mRNA expression (*in situ* hybridization) are shown. NX31975 40K PEG is shown as black, NX31976 40K PEG is shown as cross-hatched, 40K PEG is shown as white, PBS is shown as hatched, and the normal range is shown as stippled.

DETAILED DESCRIPTION OF THE INVENTION

DEFINITIONS:

"Covalent Bond" is the chemical bond formed by the sharing of electrons.

"Non-Covalent Interactions" are means by which molecular entities are held together by interactions other than Covalent Bonds including ionic interactions and hydrogen bonds.

"Lipophilic Compounds" are compounds which have the propensity to associate with or partition into lipid and/or other materials or phases with low dielectric constants, including structures that are comprised substantially of lipophilic components. Lipophilic Compounds include lipids as well as non-lipid containing compounds that have the propensity to associate with lipid (and/or other materials or phases with low dielectric constants). Cholesterol, phospholipids, and glycerolipids, such as dialkylglycerol, and diacylglycerol, and glycerol amide lipids are further examples of Lipophilic Compounds.

"Complex" as used herein describes the molecular entity formed by the covalent linking of a PDGF Nucleic Acid Ligand to a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound. In certain embodiments of the present invention, the Complex is depicted as A-B-Y, wherein A is a Lipophilic Compound or Non-Immunogenic, High Molecular Weight Compound as described herein; B is optional, and comprises a Spacer which may comprise one or more linkers Z; and Y is a PDGF Nucleic Acid Ligand.

"Lipid Constructs," for purposes of this invention, are structures containing lipids, phospholipids, or derivatives thereof comprising a variety of different structural arrangements which lipids are known to adopt in aqueous suspension. These structures include, but are not limited to, Lipid Bilayer Vesicles, micelles, Liposomes, emulsions, lipid ribbons or sheets, and may be complexed with a variety of drugs and components which are known to be pharmaceutically acceptable. In the preferred embodiment, the Lipid Construct is a Liposome. The preferred Liposome is unilamellar and has a relative size less than 200 nm. Common additional components in Lipid Constructs include cholesterol and alpha-tocopherol, among others. The Lipid Constructs may be used alone or in any combination which one skilled in the art would appreciate to provide the characteristics desired for a particular application. In addition, the technical aspects of Lipid Constructs and Liposome formation are well known in the art and any of the methods commonly practiced in the field may be used for the present invention.

"Nucleic Acid Ligand" as used herein is a non-naturally occurring Nucleic Acid having a desirable action on a Target. The Target of the present invention is PDGF, hence the term PDGF Nucleic Acid Ligand. A desirable action includes, but is not limited to, binding of the Target, catalytically changing the Target, reacting with the Target in a way which modifies/alters the Target or the functional activity of the Target, covalently attaching to the Target as in a suicide inhibitor, facilitating the reaction between the Target and another molecule. In the preferred embodiment, the action is specific binding affinity for PDGF, wherein the Nucleic Acid Ligand is not a Nucleic Acid having the known physiological function of being bound by PDGF.

In preferred embodiments of the invention, the PDGF Nucleic Acid Ligand of the Complexes and Lipid Constructs of the invention are identified by the SELEX methodology.

PDGF Nucleic Acid Ligands are identified from a Candidate Mixture of Nucleic Acids, said Nucleic Acid being a ligand of PDGF, by the method comprising a) contacting the Candidate Mixture with PDGF, wherein Nucleic Acids having an increased affinity to PDGF relative to the Candidate Mixture may be partitioned from the remainder of the Candidate Mixture; b) partitioning the increased affinity Nucleic Acids from the remainder of the Candidate Mixture; and c) amplifying the increased affinity Nucleic Acids to yield a ligand-enriched mixture of Nucleic Acids (see United States Patent Application Serial No. 08/479,725, filed June 7, 1995, entitled "High Affinity PDGF Nucleic Acid Ligands," now United States Patent No. 5,674,685, United States Patent Application Serial No. 08/479,783, filed June 7, 1995, entitled "High Affinity PDGF Nucleic Acid Ligands," now United States Patent No. 5,668,264, and United States Patent Application Serial No. 08/618,693, filed March 20, 1996, entitled "High Affinity PDGF Nucleic Acid Ligands," now U.S. Patent No. 5,723,594, which are hereby incorporated by reference herein).

In certain embodiments, portions of the PDGF Nucleic Acid Ligand (Y) may not be necessary to maintain binding and certain portions of the contiguous PDGF Nucleic Acid Ligand can be replaced with a Spacer or Linker. In these embodiments, for example, Y can be represented as Y-B'-Y'-B"-Y", wherein Y, Y' and Y" are parts of a PDGF Nucleic Acid Ligand or segments of different PDGF Nucleic Acid Ligands and B' and/or B" are Spacers or Linker molecules that replace certain nucleic acid features of the original PDGF Nucleic Acid Ligand. When B' and B" are present and Y, Y', and Y" are parts of one PDGF Nucleic Acid Ligand, a tertiary structure is formed that binds to PDGF. When B' and B" are not present, Y, Y', and Y" represent one contiguous PDGF Nucleic Acid Ligand. PDGF Nucleic Acid Ligands modified in such a manner are included in this definition.

"Candidate Mixture" is a mixture of Nucleic Acids of differing sequence from which to select a desired ligand. The source of a Candidate Mixture can be from naturally-occurring Nucleic Acids or fragments thereof, chemically synthesized Nucleic Acids, enzymatically synthesized Nucleic Acids or Nucleic Acids made by a combination of the foregoing techniques. In a preferred embodiment, each Nucleic Acid has fixed sequences surrounding a randomized region to facilitate the amplification process.

"Nucleic Acid" means either DNA, RNA, single-stranded or double-stranded and any chemical modifications thereof. Modifications include, but are not limited to, those which provide other chemical groups that incorporate additional charge, polarizability, hydrogen bonding, electrostatic interaction, and fluxionality to the Nucleic Acid Ligand bases or to the Nucleic Acid Ligand as a whole. Such modifications include, but are not limited to, 2'-position sugar modifications, 5-position pyrimidine modifications, 8-position purine modifications, modifications at exocyclic amines, substitution of 4-thiouridine, substitution of 5-bromo or 5-iodo-uracil, backbone modifications such as internucleoside phosphorothioate linkages, methylations, unusual base-pairing combinations such as the isobases isocytidine and isoguanidine and the like. Modifications can also include 3' and 5' modifications such as capping.

"Non-Immunogenic, High Molecular Weight Compound" is a compound between approximately 1000 Da to 1,000,000 Da, more preferably approximately 1000 Da to 500,000 Da, and most preferably approximately 1000 Da to 200,000 Da, that typically does not generate an immunogenic response. For the purposes of this invention, an immunogenic response is one that causes the organism to make antibody proteins. Examples of Non-Immunogenic, High Molecular Weight Compounds include Polyalkylene Glycol and polyethylene glycol. In one preferred embodiment of the invention, the Non-Immunogenic, High Molecular Weight Compound covalently linked to the PDGF Nucleic Acid Ligand is a polyalkylene glycol and has the structure $R(O(CH_2)_x)_nO-$, where R is independently selected from the group consisting of H and CH_3 , $x=2-5$, and $n \sim MW \text{ of the Polyalkylene Glycol} / (16 + 14x)$. In the preferred embodiment of the present invention, the molecular weight is about between 10-80kDa. In the most preferred embodiment, the molecular weight of the polyalkylene glycol is about between 20-45kDa. In the most preferred embodiment, $x=2$ and $n=9 \times 10^2$. There can be one or more Polyalkylene Glycols attached to the same PDGF Nucleic Acid Ligand, with the sum of the molecular weights preferably being between 10-80kDa, more preferably 20-45kDa.

In certain embodiments, the Non-Immunogenic, High Molecular Weight Compound can also be a Nucleic Acid Ligand.

"Lipid Bilayer Vesicles" are closed, fluid-filled microscopic spheres which are formed principally from individual molecules having polar (hydrophilic) and non-polar (lipophilic)

portions. The hydrophilic portions may comprise phosphato, glycerylphosphato, carboxy, sulfato, amino, hydroxy, choline and other polar groups. Examples of non-polar groups are saturated or unsaturated hydrocarbons such as alkyl, alkenyl or other lipid groups. Sterols (e.g., cholesterol) and other pharmaceutically acceptable components (including anti-oxidants like alpha-tocopherol) may also be included to improve vesicle stability or confer other desirable characteristics.

"**Liposomes**" are a subset of Lipid Bilayer Vesicles and are comprised principally of phospholipid molecules which contain two hydrophobic tails consisting of long fatty acid chains. Upon exposure to water, these molecules spontaneously align to form a bilayer membrane with the lipophilic ends of the molecules in each layer associated in the center of the membrane and the opposing polar ends forming the respective inner and outer surface of the bilayer membrane. Thus, each side of the membrane presents a hydrophilic surface while the interior of the membrane comprises a lipophilic medium. These membranes when formed are generally arranged in a system of concentric closed membranes separated by interlamellar aqueous phases, in a manner not dissimilar to the layers of an onion, around an internal aqueous space. These multilamellar vesicles (MLV) can be converted into unilamellar vesicles (UV), with the application of a shearing force.

"**Cationic Liposome**" is a Liposome that contains lipid components that have an overall positive charge at physiological pH.

"**SELEX**" methodology involves the combination of selection of Nucleic Acid Ligands which interact with a Target in a desirable manner, for example binding to a protein, with amplification of those selected Nucleic Acids. Iterative cycling of the selection/amplification steps allows selection of one or a small number of Nucleic Acids which interact most strongly with the Target from a pool which contains a very large number of Nucleic Acids. Cycling of the selection/amplification procedure is continued until a selected goal is achieved. The SELEX methodology is described in the SELEX Patent Applications.

"**Target**" means any compound or molecule of interest for which a ligand is desired. A Target can be a protein (such as PDGF, thrombin, and selectin), peptide, carbohydrate, polysaccharide, glycoprotein, hormone, receptor, antigen, antibody, virus, substrate, metabolite,

transition state analog, cofactor, inhibitor, drug, dye, nutrient, growth factor, etc. without limitation. The principal Target of the subject invention is PDGF.

"Improved Pharmacokinetic Properties" means that the PDGF Nucleic Acid Ligand covalently linked to a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound or in association with a Lipid Construct shows a longer circulation half-life *in vivo* relative to the same PDGF Nucleic Acid Ligand not in association with a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound or in association with a Lipid Construct.

"Linker" is a molecular entity that connects two or more molecular entities through Covalent Bond or Non-Covalent Interactions, and can allow spatial separation of the molecular entities in a manner that preserves the functional properties of one or more of the molecular entities. A linker can also be known as a Spacer. Examples of Linkers, include but are not limited to, the structures shown in Figures 9C-9E and the PEG spacer shown in Figure 9A.

In the preferred embodiment, the linker B' and B'' are pentaethylene glycols.

"Therapeutic" as used herein, includes treatment and/or prophylaxis. When used, Therapeutic refers to humans and other animals.

This invention includes ssDNA and RNA ligands to PDGF. This invention further includes the specific RNA ligands to PDGF shown in Tables 2-3, 6-7, and 9 and Figures 1-2, 8A and 8B (SEQ ID NOS: 4-35, 39-87, 97-149). More specifically, this invention includes nucleic acid sequences that are substantially homologous to and that have substantially the same ability to bind PDGF as the specific nucleic acid ligands shown in Tables 2-3, 6-7, and 9 and Figures 1-2, 8A and 8B (SEQ ID NOS: 4-35, 39-87, 97-149). By substantially homologous it is meant a degree of primary sequence homology in excess of 70%, most preferably in excess of 80%, and even more preferably in excess of 90%, 95%, or 99%. The percentage of homology as described herein is calculated as the percentage of nucleotides found in the smaller of the two sequences which align with identical nucleotide residues in the sequence being compared when 1 gap in a length of 10 nucleotides may be introduced to assist in that alignment.

Substantially the same ability to bind PDGF means that the affinity is within one or two orders of magnitude of the affinity of the ligands described herein. It is well within the skill of those of

ordinary skill in the art to determine whether a given sequence – substantially homologous to those specifically described herein – has the same ability to bind PDGF.

A review of the sequence homologies of the nucleic acid ligands of PDGF shown in Tables 2-3, 6-7, and 9 and Figures 1-2, 8A and 8B (SEQ ID NOS: 4-35, 39-87, 97-149) shows that sequences with little or no primary homology may have substantially the same ability to bind PDGF. For these reasons, this invention also includes Nucleic Acid Ligands that have substantially the same postulated structure or structural motifs and ability to bind PDGF as the nucleic acid ligands shown in Tables 2-3, 6-7, and 9 and Figures 1-2, 8A and 8B (SEQ ID NOS: 4-35, 39-87, 97-149). Substantially the same structure or structural motifs can be postulated by sequence alignment using the Zukerfold program (see Zuker (1989) Science 244:48-52). As would be known in the art, other computer programs can be used for predicting secondary structure and structural motifs. Substantially the same structure or structural motif of Nucleic Acid Ligands in solution or as a bound structure can also be postulated using NMR or other techniques as would be known in the art.

Further included in this invention is a method for preparing a Complex comprised of a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound by the method comprising identifying a Nucleic Acid Ligand from a Candidate Mixture of Nucleic Acids where the Nucleic Acid is a ligand of PDGF by the method of (a) contacting the Candidate Mixture of Nucleic Acids with PDGF, (b) partitioning between members of said Candidate Mixture on the basis of affinity to PDGF, and c) amplifying the selected molecules to yield a mixture of Nucleic Acids enriched for Nucleic Acid sequences with a relatively higher affinity for binding to PDGF, and covalently linking said identified PDGF Nucleic Acid Ligand with a Non-Immunogenic, High Molecular Weight Compound or a Lipophilic Compound.

It is a further object of the present invention to provide Complexes comprising one or more PDGF Nucleic Acid Ligands covalently linked to a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound. Such Complexes have one or more of the following advantages over a PDGF Nucleic Acid Ligand not in association with a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound: 1) Improved Pharmacokinetic Properties, and 2) improved capacity for intracellular delivery, or 3) improved

capacity for targeting. Complexes further associated with a Lipid Construct have the same advantages.

The Complexes or the Lipid Constructs comprising the PDGF Nucleic Acid Ligand or Complexes may benefit from one, two, or three of these advantages. For example, a Lipid Construct of the present invention may be comprised of a) a Liposome, b) a drug that is encapsulated within the interior of the Liposome, and c) a Complex comprised of a PDGF Nucleic Acid Ligand and Lipophilic Compound, wherein the PDGF Nucleic Acid Ligand component of the Complex is associated with and projecting from the exterior of the Lipid Construct. In such a case, the Lipid Construct comprising a Complex will 1) have Improved Pharmacokinetic Properties, 2) have enhanced capacity for intracellular delivery of the encapsulated drug, and 3) be specifically targeted to the preselected location in vivo that is expressing PDGF by the exteriorly associated PDGF Nucleic Acid Ligand.

In another embodiment, this invention provides a method for improving the pharmacokinetic properties of a PDGF Nucleic Acid Ligand by covalently linking the PDGF Nucleic Acid Ligand with a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound to form a Complex and administering the Complex to a patient. The invention further relates to a method for improving the pharmacokinetic properties of a PDGF Nucleic Acid Ligand by further associating the Complex with a Lipid Construct.

In another embodiment, the Complex of the present invention is comprised of a PDGF Nucleic Acid Ligand covalently attached to a Lipophilic Compound, such as a glycerolipid, or a Non-Immunogenic, High Molecular Weight Compound, such as Polyalkylene Glycol or polyethylene glycol (PEG). In these cases, the pharmacokinetic properties of the Complex will be enhanced relative to the PDGF Nucleic Acid Ligand alone. In another embodiment, the pharmacokinetic properties of the PDGF Nucleic Acid Ligand is enhanced relative to the PDGF Nucleic Acid Ligand alone when the PDGF Nucleic Acid Ligand is covalently attached to a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound and is further associated with a Lipid Construct or the PDGF Nucleic Acid Ligand is encapsulated within a Lipid Construct.

In embodiments where there are multiple PDGF Nucleic Acid Ligands, there is an increase in avidity due to multiple binding interactions with PDGF. Furthermore, in

embodiments where the Complex is comprised of multiple PDGF Nucleic Acid Ligands, the pharmacokinetic properties of the Complex will be improved relative to one PDGF Nucleic Acid Ligand alone. In embodiments where a Lipid Construct comprises multiple Nucleic Acid Ligands or Complexes, the Pharmacokinetic Properties of the PDGF Nucleic Acid Ligand may be improved relative to Lipid Constructs in which there is only one Nucleic Acid Ligand or Complex.

In certain embodiments of the invention, the Complex of the present invention is comprised of a PDGF Nucleic Acid Ligand attached to one (dimeric) or more (multimeric) other Nucleic Acid Ligands. The Nucleic Acid Ligand can be to PDGF or a different Target. In embodiments where there are multiple PDGF Nucleic Acid Ligands, there is an increase in avidity due to multiple binding interactions with PDGF. Furthermore, in embodiments of the invention where the Complex is comprised of a PDGF Nucleic Acid Ligand attached to one or more other PDGF Nucleic Acid Ligands, the pharmacokinetic properties of the Complex will be improved relative to one PDGF Nucleic Acid Ligand alone.

The Non-Immunogenic, High Molecular Weight compound or Lipophilic Compound may be covalently bound to a variety of positions on the PDGF Nucleic Acid Ligand, such as to an exocyclic amino group on the base, the 5-position of a pyrimidine nucleotide, the 8-position of a purine nucleotide, the hydroxyl group of the phosphate, or a hydroxyl group or other group at the 5' or 3' terminus of the PDGF Nucleic Acid Ligand. In embodiments where the Non-Immunogenic, High Molecular Weight Compound is polyalkylene glycol or polyethylene glycol, preferably it is bonded to the 5' or 3' hydroxyl of the phosphate group thereof. In the most preferred embodiment, the Non-Immunogenic, High Molecular Weight Compound is bonded to the 5' hydroxyl of the phosphate group of the Nucleic Acid Ligand. Attachment of the Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound to the PDGF Nucleic Acid Ligand can be done directly or with the utilization of Linkers or Spacers. In embodiments where the Lipid Construct comprises a Complex, or where the PDGF Nucleic Acid Ligands are encapsulated within the Liposome, a Non-Covalent Interaction between the PDGF Nucleic Acid Ligand or the Complex and the Lipid Construct is preferred.

One problem encountered in the therapeutic use of Nucleic Acids is that oligonucleotides in their phosphodiester form may be quickly degraded in body fluids by

intracellular and extracellular enzymes such as endonucleases and exonucleases before the desired effect is manifest. Certain chemical modifications of the PDGF Nucleic Acid Ligand can be made to increase the *in vivo* stability of the PDGF Nucleic Acid Ligand or to enhance or to mediate the delivery of the PDGF Nucleic Acid Ligand. Modifications of the PDGF Nucleic Acid Ligands contemplated in this invention include, but are not limited to, those which provide other chemical groups that incorporate additional charge, polarizability, hydrophobicity, hydrogen bonding, electrostatic interaction, and fluxionality to the PDGF Nucleic Acid Ligand bases or to the PDGF Nucleic Acid Ligand as a whole. Such modifications include, but are not limited to, 2'-position sugar modifications, 5-position pyrimidine modifications, 8-position purine modifications, modifications at exocyclic amines, substitution of 4-thiouridine, substitution of 5-bromo or 5-iodo-uracil; backbone modifications, phosphorothioate or alkyl phosphate modifications, methylations, unusual base-pairing combinations such as the isobases isocytidine and isoguanidine and the like. Modifications can also include 3' and 5' modifications such as capping.

Where the Nucleic Acid Ligands are derived by the SELEX method, the modifications can be pre- or post- SELEX modifications. Pre-SELEX modifications yield PDGF Nucleic Acid Ligands with both specificity for PDGF and improved *in vivo* stability. Post-SELEX modifications made to 2'-OH Nucleic Acid Ligands can result in improved *in vivo* stability without adversely affecting the binding capacity of the Nucleic Acid Ligands. The preferred modifications of the PDGF Nucleic Acid Ligands of the subject invention are 5' and 3' phosphorothioate capping and 3'3' inverted phosphodiester linkage at the 3' end. In the most preferred embodiment, the preferred modification of the PDGF Nucleic Acid Ligand is 3'3' inverted phosphodiester linkage at the 3' end. Additional 2' fluoro (2'-F), 2' amino (2'-NH₂) and 2' OMethyl (2'-OMe) modification of all or some of the nucleotides is preferred. In the most preferred embodiment, the preferred modification is 2'-OMe and 2'-F modification of some of the nucleotides. Additionally, the PDGF Nucleic Acid Ligand can be post-SELEX modified to substitute Linkers or Spacers such as hexaethylene glycol Spacers for certain portions.

In another aspect of the present invention, the covalent linking of the PDGF Nucleic Acid Ligand with a Non-Immunogenic, High Molecular Weight Compound or Lipophilic

Compound results in Improved Pharmacokinetic Properties (i.e., slower clearance rate) relative to the PDGF Nucleic Acid Ligand not in association with a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound.

5 In another aspect of the present invention, the Complex comprising a PDGF Nucleic Acid Ligand and Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound can be further associated with a Lipid Construct. This association may result in Improved Pharmacokinetic Properties relative to the PDGF Nucleic Acid Ligand or Complex not in association with a Lipid Construct. The PDGF Nucleic Acid Ligand or Complex can be associated with the Lipid Construct through covalent or Non-Covalent Interactions. In another
10 aspect, the PDGF Nucleic Acid Ligand can be associated with the Lipid Construct through Covalent or Non-Covalent Interactions. In a preferred embodiment, the association is through Non-Covalent Interactions. In a preferred embodiment, the Lipid Construct is a Lipid Bilayer Vesicle. In the most preferred embodiment, the Lipid Construct is a Liposome.

Liposomes for use in the present invention can be prepared by any of the various
15 techniques presently known in the art or subsequently developed. Typically, they are prepared from a phospholipid, for example, distearoyl phosphatidylcholine, and may include other materials such as neutral lipids, for example, cholesterol, and also surface modifiers such as positively charged (e.g., sterylamine or aminomannose or aminomannitol derivatives of cholesterol) or negatively charged (e.g., diacetyl phosphate, phosphatidyl glycerol) compounds.
20 Multilamellar Liposomes can be formed by conventional techniques, that is, by depositing a selected lipid on the inside wall of a suitable container or vessel by dissolving the lipid in an appropriate solvent, and then evaporating the solvent to leave a thin film on the inside of the vessel or by spray drying. An aqueous phase is then added to the vessel with a swirling or vortexing motion which results in the formation of MLVs. UVs can then be formed by
25 homogenization, sonication or extrusion (through filters) of MLV's. In addition, UVs can be formed by detergent removal techniques.

In certain embodiments of this invention, the Lipid Construct comprises a targeting PDGF Nucleic Acid Ligand(s) associated with the surface of the Lipid Construct and an encapsulated therapeutic or diagnostic agent. Preferably the Lipid Construct is a Liposome.
30 Preformed Liposomes can be modified to associate with the PDGF Nucleic Acid Ligands.

For example, a Cationic Liposome associates through electrostatic interactions with the PDGF Nucleic Acid Ligand. A PDGF Nucleic Acid Ligand covalently linked to a Lipophilic Compound, such as a glycerolipid, can be added to preformed Liposomes whereby the glycerolipid, phospholipid, or glycerol amide lipid becomes associated with the liposomal membrane. Alternatively, the PDGF Nucleic Acid Ligand can be associated with the Liposome during the formulation of the Liposome.

It is well known in the art that Liposomes are advantageous for encapsulating or incorporating a wide variety of therapeutic and diagnostic agents. Any variety of compounds can be enclosed in the internal aqueous compartment of the Liposomes. Illustrative therapeutic agents include antibiotics, antiviral nucleosides, antifungal nucleosides, metabolic regulators, immune modulators, chemotherapeutic drugs, toxin antidotes, DNA, RNA, antisense oligonucleotides, etc. By the same token, the Lipid Bilayer Vesicles may be loaded with a diagnostic radionuclide (e.g., Indium 111, Iodine 131, Yttrium 90, Phosphorous 32, or gadolinium) and fluorescent materials or other materials that are detectable in *in vitro* and *in vivo* applications. It is to be understood that the therapeutic or diagnostic agent can be encapsulated by the Liposome walls in the aqueous interior. Alternatively, the carried agent can be a part of, that is, dispersed or dissolved in the vesicle wall-forming materials.

During Liposome formation, water soluble carrier agents may be encapsulated in the aqueous interior by including them in the hydrating solution, and lipophilic molecules incorporated into the lipid bilayer by inclusion in the lipid formulation. In the case of certain molecules (e.g., cationic or anionic lipophilic drugs), loading of the drug into preformed Liposomes may be accomplished, for example, by the methods described in U.S. Patent No. 4,946,683, the disclosure of which is incorporated herein by reference. Following drug encapsulation, the Liposomes are processed to remove unencapsulated drug through processes such as gel chromatography or ultrafiltration. The Liposomes are then typically sterile filtered to remove any microorganisms which may be present in the suspension. Microorganisms may also be removed through aseptic processing.

If one wishes to encapsulate large hydrophilic molecules with Liposomes, larger unilamellar vesicles can be formed by methods such as the reverse-phase evaporation (REV) or solvent infusion methods. Other standard methods for the formation of Liposomes are

known in the art, for example, methods for the commercial production of Liposomes include the homogenization procedure described in U.S. Patent No. 4,753,788 and the thin-film evaporation method described in U.S. Patent No. 4,935,171, which are incorporated herein by reference.

5 It is to be understood that the therapeutic or diagnostic agent can also be associated with the surface of the Lipid Bilayer Vesicle. For example, a drug can be attached to a phospholipid or glyceride (a prodrug). The phospholipid or glyceride portion of the prodrug can be incorporated into the lipid bilayer of the Liposome by inclusion in the lipid formulation or loading into preformed Liposomes (see U.S. Patent Nos 5,194,654 and
10 5,223,263, which are incorporated by reference herein).

 It is readily apparent to one skilled in the art that the particular Liposome preparation method will depend on the intended use and the type of lipids used to form the bilayer membrane.

 Lee and Low (1994, JBC 269:3198-3204) and DeFrees *et al.* (1996, JACS 118:6101-
15 6104) first showed that co-formulation of ligand-PEG-lipid with lipid components gave liposomes with both inward and outward facing orientations of the PEG-ligand. Passive anchoring was outlined by Zalipsky *et al.* (1997, Bioconj. Chem. 8:111-118) as a method for anchoring oligopeptide and oligosaccharide ligands exclusively to the external surface of liposomes. The central concept presented in their work is that oligo-PEG-lipid conjugates can
20 be prepared and then formulated into pre-formed liposomes via spontaneous incorporation (“anchoring”) of the lipid tail into the existing lipid bilayer. The lipid group undergoes this insertion in order to reach a lower free energy state via the removal of its hydrophobic lipid anchor from aqueous solution and its subsequent positioning in the hydrophobic lipid bilayer. The key advantage to such a system is that the oligo-lipid is anchored exclusively to the
25 exterior of the lipid bilayer. Thus, no oligo-lipids are wasted by being unavailable for interactions with their biological targets by being in an inward-facing orientation.

 The efficiency of delivery of a PDGF Nucleic Acid Ligand to cells may be optimized by using lipid formulations and conditions known to enhance fusion of Liposomes with cellular membranes. For example, certain negatively charged lipids such as
30 phosphatidylglycerol and phosphatidylserine promote fusion, especially in the presence of

other fusogens (e.g., multivalent cations like Ca^{2+} , free fatty acids, viral fusion proteins, short chain PEG, lysolecithin, detergents and surfactants). Phosphatidylethanolamine may also be included in the Liposome formulation to increase membrane fusion and, concomitantly, enhance cellular delivery. In addition, free fatty acids and derivatives thereof, containing, for example, carboxylate moieties, may be used to prepare pH-sensitive Liposomes which are negatively charged at higher pH and neutral or protonated at lower pH. Such pH-sensitive Liposomes are known to possess a greater tendency to fuse.

In the preferred embodiment, the PDGF Nucleic Acid Ligands of the present invention are derived from the SELEX methodology. SELEX is described in U.S. Patent Application Serial No. 07/536,428, entitled Systematic Evolution of Ligands by Exponential Enrichment, now abandoned, U.S. Patent Application Serial No. 07/714,131, filed June 10, 1991, entitled Nucleic Acid Ligands, now United States Patent No. 5,475,096, and United States Patent Application Serial No. 07/931,473, filed August 17, 1992, entitled "Methods for Identifying Nucleic Acid Ligands," now United States Patent No. 5,270,163 (see also WO 91/19813).

These applications, each specifically incorporated herein by reference, are collectively called the SELEX Patent Applications.

The SELEX process provides a class of products which are Nucleic Acid molecules, each having a unique sequence, and each of which has the property of binding specifically to a desired Target compound or molecule. Target molecules are preferably proteins, but can also include among others carbohydrates, peptidoglycans and a variety of small molecules. SELEX methodology can also be used to Target biological structures, such as cell surfaces or viruses, through specific interaction with a molecule that is an integral part of that biological structure.

In its most basic form, the SELEX process may be defined by the following series of steps:

- 1) A Candidate Mixture of Nucleic Acids of differing sequence is prepared. The Candidate Mixture generally includes regions of fixed sequences (i.e., each of the members of the Candidate Mixture contains the same sequences in the same location) and regions of randomized sequences. The fixed sequence regions are selected either: (a) to assist in the amplification steps described below, (b) to mimic a sequence known to bind to the Target, or (c) to enhance the concentration of a given structural arrangement of the Nucleic Acids in the

Candidate Mixture. The randomized sequences can be totally randomized (i.e., the probability of finding a base at any position being one in four) or only partially randomized (e.g., the probability of finding a base at any location can be selected at any level between 0 and 100 percent).

2) The Candidate Mixture is contacted with the selected Target under conditions favorable for binding between the Target and members of the Candidate Mixture. Under these circumstances, the interaction between the Target and the Nucleic Acids of the Candidate Mixture can be considered as forming Nucleic Acid-target pairs between the Target and those Nucleic Acids having the strongest affinity for the Target.

3) The Nucleic Acids with the highest affinity for the target are partitioned from those Nucleic Acids with lesser affinity to the target. Because only an extremely small number of sequences (and possibly only one molecule of Nucleic Acid) corresponding to the highest affinity Nucleic Acids exist in the Candidate Mixture, it is generally desirable to set the partitioning criteria so that a significant amount of the Nucleic Acids in the Candidate Mixture (approximately 5-50%) are retained during partitioning.

4) Those Nucleic Acids selected during partitioning as having the relatively higher affinity for the target are then amplified to create a new Candidate Mixture that is enriched in Nucleic Acids having a relatively higher affinity for the target.

5) By repeating the partitioning and amplifying steps above, the newly formed Candidate Mixture contains fewer and fewer unique sequences, and the average degree of affinity of the Nucleic Acids to the target will generally increase. Taken to its extreme, the SELEX process will yield a Candidate Mixture containing one or a small number of unique Nucleic Acids representing those Nucleic Acids from the original Candidate Mixture having the highest affinity to the target molecule.

The basic SELEX method has been modified to achieve a number of specific objectives. For example, United States Patent Application Serial No. 07/960,093, filed October 14, 1992, entitled "Method for Selecting Nucleic Acids on the Basis of Structure," describes the use of SELEX in conjunction with gel electrophoresis to select Nucleic Acid molecules with specific structural characteristics, such as bent DNA. United States Patent Application Serial No. 08/123,935, filed September 17, 1993, entitled "Photoselection of Nucleic Acid Ligands,"

describes a SELEX based method for selecting Nucleic Acid Ligands containing photoreactive groups capable of binding and/or photocrosslinking to and/or photoinactivating a target molecule. United States Patent Application Serial No. 08/134,028, filed October 7, 1993, entitled "High-Affinity Nucleic Acid Ligands That Discriminate Between Theophylline and Caffeine," now abandoned (see United States Patent No. 5,580,737), describes a method for identifying highly specific Nucleic Acid Ligands able to discriminate between closely related molecules, termed Counter-SELEX. United States Patent Application Serial No. 08/143,564, filed October 25, 1993, entitled "Systematic Evolution of Ligands by Exponential Enrichment: Solution SELEX," now abandoned (see United States Patent No. 5,567,588), describes a SELEX-based method which achieves highly efficient partitioning between oligonucleotides having high and low affinity for a target molecule. United States Patent Application Serial No. 07/964,624, filed October 21, 1992, entitled "Nucleic Acid Ligands to HIV-RT and HIV-1 Rev," now United States Patent No. 5,496,938, describes methods for obtaining improved Nucleic Acid Ligands after SELEX has been performed. United States Patent Application Serial No. 08/400,440, filed March 8, 1995, entitled "Systematic Evolution of Ligands by Exponential Enrichment: Chemi-SELEX," now U.S. Patent No. 5,705,337, describes methods for covalently linking a ligand to its target.

The SELEX method encompasses the identification of high-affinity Nucleic Acid Ligands containing modified nucleotides conferring improved characteristics on the ligand, such as improved *in vivo* stability or improved delivery characteristics. Examples of such modifications include chemical substitutions at the ribose and/or phosphate and/or base positions. SELEX-identified Nucleic Acid Ligands containing modified nucleotides are described in United States Patent Application Serial No. 08/117,991, filed September 8, 1993, entitled "High Affinity Nucleic Acid Ligands Containing Modified Nucleotides," now United States Patent No. 5,660,985, that describes oligonucleotides containing nucleotide derivatives chemically modified at the 5- and 2'-positions of pyrimidines. United States Patent Application Serial No. 08/134,028, *supra*, describes highly specific Nucleic Acid Ligands containing one or more nucleotides modified with 2'-amino (2'-NH₂), 2'-fluoro (2'-F), and/or 2'-O-methyl (2'-OMe). United States Patent Application Serial No. 08/264,029, filed June 22, 1994, entitled "Novel Method of Preparation of Known and Novel 2' Modified Nucleosides by Intramolecular

Nucleophilic Displacement," describes oligonucleotides containing various 2'-modified pyrimidines.

The SELEX method encompasses combining selected oligonucleotides with other selected oligonucleotides and non-oligonucleotide functional units as described in United States Patent Application Serial No. 08/284,063, filed August 2, 1994, entitled "Systematic Evolution of Ligands by Exponential Enrichment: Chimeric SELEX," now United States Patent No. 5,637,459, and United States Patent Application Serial No. 08/234,997, filed April 28, 1994, entitled "Systematic Evolution of Ligands by Exponential Enrichment: Blended SELEX," now U.S. Patent No. 5,683,867, respectively. These applications allow the combination of the broad array of shapes and other properties, and the efficient amplification and replication properties, of oligonucleotides with the desirable properties of other molecules.

The SELEX method further encompasses combining selected Nucleic Acid Ligands with Lipophilic Compounds or Non-Immunogenic, High Molecular Weight Compounds in a diagnostic or therapeutic Complex as described in United States Patent Application Serial No. 08/434,465, filed May 4, 1995, entitled Nucleic Acid Ligand Complexes," now U.S. Patent No. 6,011,021. The SELEX method further encompasses combining selected VEGF Nucleic Acid Ligands with lipophilic compounds, such as diacyl glycerol or dialkyl glycerol, as described in United States Patent Application Serial No. 08/739,109, filed October 25, 1996, entitled "Vascular Endothelial Growth Factor (VEGF) Nucleic Acid Ligand Complexes," now U.S. Patent No. 5,859,228. VEGF Nucleic Acid Ligands that are associated with a High Molecular Weight, Non-Immunogenic Compound, such as Polyethyleneglycol, or a Lipophilic Compound, such as Glycerolipid, phospholipid, or glycerol amide lipid, in a diagnostic or therapeutic complex are described in United States Patent Application Serial No. 08/897,351, filed July 21, 1997, entitled "Vascular Endothelial Growth Factor (VEGF) Nucleic Acid Ligand Complexes," now U.S. Patent No. 6,051,698. Each of the above described patent applications which describe modifications of the basic SELEX procedure are specifically incorporated by reference herein in their entirety.

SELEX identifies Nucleic Acid Ligands that are able to bind targets with high affinity and with outstanding specificity, which represents a singular achievement that is unprecedented

in the field of Nucleic Acids research. These characteristics are, of course, the desired properties one skilled in the art would seek in a therapeutic or diagnostic ligand.

In order to produce Nucleic Acid Ligands desirable for use as a pharmaceutical, it is preferred that the Nucleic Acid Ligand (1) binds to the target in a manner capable of achieving the desired effect on the target; (2) be as small as possible to obtain the desired effect; (3) be as stable as possible; and (4) be a specific ligand to the chosen target. In most situations, it is preferred that the Nucleic Acid Ligand has the highest possible affinity to the target. Additionally, Nucleic Acid Ligands can have facilitating properties.

In commonly assigned U.S. Patent Application Serial No. 07/964,624, filed October 21, 1992 ('624), now United States Patent No. 5,496,938, methods are described for obtaining improved Nucleic Acid Ligands after SELEX has been performed. The '624 application, entitled Nucleic Acid Ligands to HIV-RT and HIV-1 Rev, is specifically incorporated herein by reference.

The SELEX process has been used to identify a group of high affinity RNA Ligands to PDGF from random ssDNA libraries and 2'-fluoro-2'-deoxypyrimidine RNA ligands from random ssDNA libraries (United States Patent Application Serial No. 08/618,693, filed March 20, 1996, entitled High-Affinity PDGF Nucleic Acid Ligands, now U.S. Patent No. 5,723,594, which is a Continuation-in-Part Application of United States Patent Application Serial No. 08/479,783, filed June 7, 1995, entitled High-Affinity PDGF Nucleic Acid Ligands, now U.S. Patent No. 5,668,264, and United States Patent Application Serial No. 08/479,725, filed June 7, 1995, entitled "High Affinity PDGF Nucleic Acid Ligands, now U.S. Patent No. 5,674,685, both of which are incorporated herein by reference; see also Green *et al.* (1995) Chemistry and Biology 2:683-695).

In embodiments where the PDGF Nucleic Acid Ligand(s) can serve in a targeting capacity, the PDGF Nucleic Acid Ligands adopt a three dimensional structure that must be retained in order for the PDGF Nucleic Acid Ligand to be able to bind its target. In embodiments where the Lipid Construct comprises a Complex and the PDGF Nucleic Acid Ligand of the Complex is projecting from the surface of the Lipid Construct, the PDGF Nucleic Acid Ligand must be properly oriented with respect to the surface of the Lipid Construct so that its target binding capacity is not compromised. This can be accomplished by attaching the

PDGF Nucleic Acid Ligand at a position that is distant from the binding portion of the PDGF Nucleic Acid Ligand. The three dimensional structure and proper orientation can also be preserved by use of a Linker or Spacer as described supra.

Any variety of therapeutic or diagnostic agents can be attached to the Complex for
5 targeted delivery by the Complex. In addition, any variety of therapeutic or diagnostic agents can be attached encapsulated, or incorporated into the Lipid Construct as discussed supra for targeted delivery by the Lipid Construct.

In embodiments where the Complex is comprised of a Lipophilic Compound and a PDGF Nucleic Acid Ligand in association with a Liposome, for example, the PDGF Nucleic
10 Acid Ligand could target tumor cells expressing PDGF(e.g., in Kaposi's sarcoma) for delivery of an antitumor drug (e.g., daunorubicin) or imaging agent (e.g., radiolabels). It should be noted that cells and tissues surrounding the tumor may also express PDGF, and targeted delivery of an antitumor drug to these cells would also be effective.

In an alternative embodiment, the therapeutic or diagnostic agent to be delivered to
15 the Target cell could be another Nucleic Acid Ligand.

It is further contemplated by this invention that the agent to be delivered can be incorporated into the Complex in such a way as to be associated with the outside surface of the Liposome (e.g., a prodrug, receptor antagonist, or radioactive substance for treatment or
imaging). As with the PDGF Nucleic Acid Ligand, the agent can be associated through
20 covalent or Non-Covalent Interactions. The Liposome would provide targeted delivery of the agent extracellularly, with the Liposome serving as a Linker.

In another embodiment, a Non-Immunogenic, High Molecular Weight Compound (e.g., PEG) can be attached to the Liposome to provide Improved Pharmacokinetic Properties for the Complex. PDGF Nucleic Acid Ligands may be attached to the Liposome membrane
25 or may be attached to a Non-Immunogenic, High Molecular Weight Compound which in turn is attached to the membrane. In this way, the Complex may be shielded from blood proteins and thus be made to circulate for extended periods of time while the PDGF Nucleic Acid Ligand is still sufficiently exposed to make contact with and bind to its Target.

In another embodiment of the present invention, more than one PDGF Nucleic Acid
30 Ligand is attached to the surface of the same Liposome. This provides the possibility of

bringing the same PDGF molecules in close proximity to each other and can be used to generate specific interactions between the PDGF molecules.

In an alternative embodiment of the present invention, PDGF Nucleic Acid Ligands and a Nucleic Acid Ligand to a different Target can be attached to the surface of the same
5 Liposome. This provides the possibility of bringing PDGF in close proximity to a different Target and can be used to generate specific interactions between PDGF and the other Target. In addition to using the Liposome as a way of bringing Targets in close proximity, agents could be encapsulated in the Liposome to increase the intensity of the interaction.

The Lipid Construct comprising a Complex allows for the possibility of multiple
10 binding interactions to PDGF. This, of course, depends on the number of PDGF Nucleic Acid Ligands per Complex, and the number of Complexes per Lipid Construct, and mobility of the PDGF Nucleic Acid Ligands and receptors in their respective membranes. Since the effective binding constant may increase as the product of the binding constant for each site, there is a substantial advantage to having multiple binding interactions. In other words, by
15 having many PDGF Nucleic Acid Ligands attached to the Lipid Construct, and therefore creating multivalency, the effective affinity (i.e., the avidity) of the multimeric Complex for its Target may become as good as the product of the binding constant for each site.

In certain embodiments of the invention, the Complex of the present invention is comprised of a PDGF Nucleic Acid Ligand attached to a Lipophilic Compound. In this case,
20 the pharmacokinetic properties of the Complex will be improved relative to the PDGF Nucleic Acid Ligand alone. As discussed *supra*, the Lipophilic Compound may be covalently bound to the PDGF Nucleic Acid Ligand at numerous positions on the PDGF Nucleic Acid Ligand.

In another embodiment of the invention, the Lipid Construct comprises a PDGF
25 Nucleic Acid Ligand or Complex. In this embodiment, the glycerolipid can assist in the incorporation of the PDGF Nucleic Acid Ligand into the Liposome due to the propensity for a glycerolipid to associate with other Lipophilic Compounds. The glycerolipid in association with a PDGF Nucleic Acid Ligand can be incorporated into the lipid bilayer of the Liposome by inclusion in the formulation or by loading into preformed Liposomes. The glycerolipid
30 can associate with the membrane of the Liposome in such a way so as the PDGF Nucleic

Acid Ligand is projecting into or out of the Liposome. In embodiments where the PDGF Nucleic Acid Ligand is projecting out of the Complex, the PDGF Nucleic Acid Ligand can serve in a targeting capacity. It is to be understood that additional compounds can be associated with the Lipid Construct to further improve the Pharmacokinetic Properties of the Lipid Construct. For example, a PEG may be attached to the exterior-facing part of the membrane of the Lipid Construct.

In other embodiments, the Complex of the present invention is comprised of a PDGF Nucleic Acid Ligand covalently linked to a Non-Immunogenic, High Molecular Weight Compound such as Polyalkylene Glycol or PEG. In this embodiment, the pharmacokinetic properties of the Complex are improved relative to the PDGF Nucleic Acid Ligand alone. The Polyalkylene Glycol or PEG may be covalently bound to a variety of positions on the PDGF Nucleic Acid Ligand. In embodiments where Polyalkylene Glycol or PEG are used, it is preferred that the PDGF Nucleic Acid Ligand is bonded through the 5' hydroxyl group via a phosphodiester linkage.

In certain embodiments, a plurality of Nucleic Acid Ligands can be associated with a single Non-Immunogenic, High Molecular Weight Compound, such as Polyalkylene Glycol or PEG, or a Lipophilic Compound, such as a glycerolipid. The Nucleic Acid Ligands can all be to PDGF or PDGF and a different Target. In embodiments where there are multiple PDGF Nucleic Acid Ligands, there is an increase in avidity due to multiple binding interactions with PDGF. In yet further embodiments, a plurality of Polyalkylene Glycol, PEG, glycerol lipid molecules can be attached to each other. In these embodiments, one or more PDGF Nucleic Acid Ligands or Nucleic Acid Ligands to PDGF and other Targets can be associated with each Polyalkylene Glycol, PEG, or glycerol lipid. This also results in an increase in avidity of each Nucleic Acid Ligand to its Target. In embodiments where multiple PDGF Nucleic Acid Ligands are attached to Polyalkylene Glycol, PEG, or glycerol lipid, there is the possibility of bringing PDGF molecules in close proximity to each other in order to generate specific interactions between PDGF. Where multiple Nucleic Acid Ligands specific for PDGF and different Targets are attached to Polyalkylene Glycol, PEG, or glycerol lipid, there is the possibility of bringing PDGF and another Target in close proximity to each other in order to generate specific interactions between the PDGF and the other Target. In addition, in

embodiments where there are Nucleic Acid Ligands to PDGF or Nucleic Acid Ligands to PDGF and different Targets associated with Polyalkylene Glycol, PEG, or glycerol lipid, a drug can also be associated with Polyalkylene Glycol, PEG, or glycerol lipid. Thus the Complex would provide targeted delivery of the drug, with Polyalkylene Glycol, PEG, or glycerol lipid serving as a Linker.

PDGF Nucleic Acid Ligands selectively bind PDGF. Thus, a Complex comprising a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound or a Lipid Construct comprising a PDGF Nucleic Acid Ligand or a Complex are useful as pharmaceuticals or diagnostic agents. The PDGF Nucleic Acid Ligand-containing Complexes and Lipid Constructs can be used to treat, inhibit, prevent or diagnose any disease state that involves inappropriate PDGF production, for example, cancer, angiogenesis, restenosis, and fibrosis. PDGF is produced and secreted in varying amounts by many tumor cells. Thus, the present invention, includes methods of treating, inhibiting, preventing, or diagnosing cancer by administration of a Complex comprising a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound, a Lipid Construct comprising a Complex, or a PDGF Nucleic Acid Ligand in association with a Lipid Construct without being part of the Complex.

Angiogenesis rarely occurs in healthy adults, except during the menstrual cycle and wound healing. Angiogenesis is a central feature, however, of various disease states, including, but not limited to cancer, diabetic retinopathy, macular degeneration, psoriasis and rheumatoid arthritis. The present invention, therefore, includes methods of treating, inhibiting, preventing, or diagnosing angiogenesis by administration of a Complex comprising PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound, a Lipid Construct comprising PDGF Nucleic Acid Ligand or a Complex comprising a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound.

PDGF is also produced in fibrosis in organs, such as lung, bone marrow and kidney. Fibrosis can also be associated with radiation treatments. The present invention, therefore, includes methods of treating, inhibiting, preventing or diagnosing lung, bone marrow, kidney and radiation treatment-associated fibrosis by administration of a Complex comprising PDGF

Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound, a Lipid Construct comprising PDGF Nucleic Acid Ligand or a Complex comprising a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound.

5 PDGF is a prominent growth factor involved in restenosis. Restenosis, the reocclusion of a diseased blood vessel after treatment to eliminate stenosis, is a common occurrence that develops following coronary interventions and some peripheral vessel interventions. Additionally, stents have been used in the treatment of or in conjunction with treatment of coronary and non-coronary vessels; however, restenosis is also associated with use of stents
10 (called in-stent restenosis). In-stent restenosis occurs in about 15-30% of coronary interventions and frequently in some peripheral vessel interventions. For example, in-stent restenosis is a significant problem in small vessels, with frequencies ranging from 15% to 40% in stented femoral or popliteal arteries. Intermediate-sized vessels, such as renal arteries, have an in-stent restenosis rate of 10-20%.

15 The present invention, therefore, includes methods of treating, inhibiting, preventing or diagnosing restenosis by administration of a Complex comprising PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound, a Lipid Construct comprising PDGF Nucleic Acid Ligand or a Complex comprising a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or
20 Lipophilic Compound. The present invention also includes methods of treating, inhibiting, preventing or diagnosing restenosis in coronary and non-coronary vessels. The present invention also includes methods of treating, inhibiting, preventing or diagnosing in-stent restenosis.

25 Additionally, cancer, angiogenesis, restenosis, and fibrosis involve the production of growth factors other than PDGF. Thus, it is contemplated by this invention that a Complex comprising PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound, a Lipid Construct comprising PDGF Nucleic Acid Ligand or a Complex comprising a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound can be used in conjunction with
30 Complexes comprising Nucleic Acid Ligands to other growth factors (such as bFGF,

TGF α , hKGF, etc.) and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound, a Lipid Construct comprising PDGF Nucleic Acid Ligand or a Complex comprising a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound.

5 In one embodiment of the present invention, the Lipid Construct comprises a Complex comprised of a PDGF Nucleic Acid Ligand and a Lipophilic Compound with an additional diagnostic or therapeutic agent encapsulated in the Lipid Construct or associated with the interior of the Lipid Construct. In the preferred embodiment, the Lipid Construct is a Lipid Bilayer Vesicle, and more preferably a Liposome. The therapeutic use of Liposomes includes
10 the delivery of drugs which are normally toxic in the free form. In the liposomal form, the toxic drug is occluded, and may be directed away from the tissues sensitive to the drug and targeted to selected areas. Liposomes can also be used therapeutically to release drugs over a prolonged period of time, reducing the frequency of administration. In addition, liposomes can provide a method for forming aqueous dispersions of hydrophobic or amphiphilic drugs, which are
15 normally unsuitable for intravenous delivery.

In order for many drugs and imaging agents to have therapeutic or diagnostic potential, it is necessary for them to be delivered to the proper location in the body, and the liposome can thus be readily injected and form the basis for sustained release and drug delivery to specific cell types, or parts of the body. Several techniques can be employed to use liposomes to target
20 encapsulated drugs to selected host tissues, and away from sensitive tissues. These techniques include manipulating the size of the liposomes, their net surface charge, and their route of administration. MLVs, primarily because they are relatively large, are usually rapidly taken up by the reticuloendothelial system (principally the liver and spleen). UVs, on the other hand, have been found to exhibit increased circulation times, decreased clearance rates and greater
25 biodistribution relative to MLVs.

Passive delivery of liposomes involves the use of various routes of administration, e.g., intravenous, subcutaneous, intramuscular and topical. Each route produces differences in localization of the liposomes. Two common methods used to direct liposomes actively to selected target areas involve attachment of either antibodies or specific receptor ligands to the
30 surface of the liposomes. In one embodiment of the present invention, the PDGF Nucleic Acid

Ligand is associated with the outside surface of the liposome, and serves in a targeting capacity. Additional targeting components, such as antibodies or specific receptor ligands can be included on the liposome surface, as would be known to one of skill in the art. In addition, some efforts have been successful in targeting liposomes to tumors without the use of antibodies, see, for example, U.S. Patent No. 5,019,369, U.S. Patent No. 5,435,989, and U.S. Patent No. 4,441,775, and it would be known to one of skill in the art to incorporate these alternative targeting methods.

Therapeutic or diagnostic compositions of a Complex comprising PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound, a Lipid Construct comprising a Complex comprised of a PDGF Nucleic Acid Ligand and a Non-Immunogenic, High Molecular Weight Compound or Lipophilic Compound, and a PDGF Nucleic Acid Ligand in association with a Lipid Construct without being part of a Complex may be administered parenterally by injection, although other effective administration forms, such as intraarticular injection, inhalant mists, orally active formulations, transdermal iotophoresis or suppositories, are also envisioned. They may also be applied locally by direct injection, can be released from devices, such as implanted stents or catheters, or delivered directly to the site by an infusion pump. One preferred carrier is physiological saline solution, but it is contemplated that other pharmaceutically acceptable carriers may also be used. In one embodiment, it is envisioned that the carrier and the PDGF Nucleic Acid Ligand Complex constitute a physiologically-compatible, slow release formulation. The primary solvent in such a carrier may be either aqueous or non-aqueous in nature. In addition, the carrier may contain other pharmacologically-acceptable excipients for modifying or maintaining the pH, osmolarity, viscosity, clarity, color, sterility, stability, rate of dissolution, or odor of the formulation. Similarly, the carrier may contain still other pharmacologically-acceptable excipients for modifying or maintaining the stability, rate of dissolution, release, or absorption of the PDGF Nucleic Acid Ligand. Such excipients are those substances usually and customarily employed to formulate dosages for parental administration in either unit dose or multi-dose form.

Once the therapeutic or diagnostic composition has been formulated, it may be stored in sterile vials as a solution, suspension, gel, emulsion, solid, or dehydrated or lyophilized powder.

Such formulations may be stored either in ready to use form or requiring reconstitution immediately prior to administration. The manner of administering formulations containing PDGF Nucleic Acid Ligand for systemic delivery may be via subcutaneous, intramuscular, intravenous, intranasal or vaginal or rectal suppository.

5 The advantages of the Complexes and Lipid Constructs of the invention include: i) improving the plasma pharmacokinetics of the Nucleic Acid Ligand; ii) Presenting Nucleic Acid Ligands in a multivalent array with the aim of increasing the avidity of interaction with their targets; iii) combining two or more presenting Nucleic Acid Ligands with different specificities in the same liposome particle; iv) enhancing the delivery of presenting Nucleic
10 Acid Ligands to tumors by taking advantage of the intrinsic tumor targeting properties of liposomes; and v) using the high affinity and specificity of presenting Nucleic Acid Ligands, which is comparable to that of antibodies, to guide liposomal contents to specific targets. presenting Nucleic Acid Ligands are well suited for the kinds of preparations described here since, unlike most proteins, the denaturation of presenting Nucleic Acid Ligands by heat,
15 various molecular denaturants and organic solvents is readily reversible.

 The following examples are provided to explain and illustrate the present invention and are not to be taken as limiting of the invention. Example 1 describes the various materials and experimental procedures used in Examples 2-4 for the generation of ssDNA ligands to PDGF and tests associated therewith. Example 2 describes the ssDNA ligands to PDGF and the
20 predicted secondary structure of selected nucleic acid ligands and a shared secondary structure motif. Example 3 describes the minimum sequence necessary for high affinity binding, the sites on the nucleic acid ligands and PDGF that are in contact, inhibition by DNA ligands of PDGF isoforms on cultured cells, and inhibition of mitogenic effects of PDGF in cells by DNA ligands. Example 4 describes substitutions of SELEX-derived ligands with modified
25 nucleotides. Example 5 describes synthesis of PEG-modified PDGF Nucleic Acid Ligands. Example 6 describes stability of modified ligands in serum. Example 7 describes efficacy of a modified ligand (NX31975-40K PEG) in restenosis. Example 8 describes the various materials and method used in Example 9 for testing the inhibition of PDGF in glomerulonephritis. Example 9 describes inhibition of PDGF in glomerulonephritis. Example 10 describes the

experimental procedures for evolving 2'-fluoro-2'-deoxypyrimidine RNA ligands to PDGF and the RNA sequences obtained.

EXAMPLE 1. EXPERIMENTAL PROCEDURES

This example provides the general procedures followed and incorporated in Examples 2-4.

MATERIALS.

Recombinant human PDGF-AA (Mr=29,000), PDGF-AB (Mr=27,000) and PDGF-BB (Mr=25,000) were purchased from R&D Systems (Minneapolis, MN) in lyophilized form, free from carrier protein. All three isoforms were produced in *E. coli* from synthetic genes based on the sequences for the long form of the mature human PDGF A-chain (Betsholtz *et al.* (1986) Nature 320:695-699) and the naturally occurring mature form of human PDGF B-chain (Johnsson *et al.* (1984) EMBO J. 3:921-928). Randomized DNA libraries, PCR primers and DNA ligands and 5'-iodo-2'-deoxyuridine-substituted DNA ligands were synthesized by NeXstar Pharmaceuticals, Inc. (Boulder, CO) or by Operon Technologies (Alameda, CA) using the standard solid phase phosphoramidite method (Sinha *et al.* (1984) Nucleic Acids Res. 12:4539-4557).

SINGLE STRANDED DNA (ssDNA) SELEX

Essential features of the SELEX procedure have been described in detail in the SELEX Patent Applications (see also Tuerk and Gold (1990) Science 249:505; Jellinek *et al.* (1994) Biochemistry 33:10450; Jellinek *et al.* (1993) Proc. Natl. Acad. Sci. USA 90:1227), which are incorporated by reference herein. The initial ssDNA library containing a contiguous randomized region of forty nucleotides, flanked by primer annealing regions (Table 1) (SEQ ID NOS: 1-3) of invariant sequence, was synthesized by the solid phase phosphoramidite method using equal molar mixture of the four phosphoramidites to generate the randomized positions. The ssDNA library was purified by electrophoresis on an 8% polyacrylamide/7 M urea gel. The band that corresponds to the full-length DNA was visualized under UV light, excised from the gel, eluted by the crush and soak method, ethanol precipitated and pelleted by centrifugation. The pellet was dried under vacuum and

resuspended in phosphate buffered saline supplemented with 1 mM MgCl₂ (PBSM = 10.1 mM Na₂HPO₄, 1.8 mM KH₂PO₄, 137 mM NaCl and 2.7 mM KCl, 1 mM MgCl₂, pH 7.4) buffer. Prior to incubation with the protein, the ssDNA was heated at 90°C for 2 minutes in PBSM and cooled on ice. The first selection was initiated by incubating approximately 500 pmol (3 x 10¹⁴ molecules) of 5' ³²P end-labeled random ssDNA with PDGF-AB in binding buffer (PBSM containing 0.01% human serum albumin (HSA)). The mixture was incubated at 4°C overnight, followed by a brief (15 min) incubation at 37°C. The DNA bound to PDGF-AB was separated from unbound DNA by electrophoresis on an 8% polyacrylamide gel (1:30 bis-acrylamide:acrylamide) at 4°C and at 5 V/cm with 89 mM Tris-borate (pH 8.3) containing 2 mM EDTA as the running buffer. The band that corresponds to the PDGF-ssDNA complex, which runs with about half the electrophoretic mobility of the free ssDNA, was visualized by autoradiography, excised from the gel and eluted by the crush and soak method. In subsequent affinity selections, the ssDNA was incubated with PDGF-AB for 15 minutes at 37°C in binding buffer and the PDGF-bound ssDNA was separated from the unbound DNA by nitrocellulose filtration, as previously described (Green *et al.* (1995) Chemistry and Biology 2:683-695). All affinity-selected ssDNA pools were amplified by PCR in which the DNA was subjected to 12-20 rounds of thermal cycling (30 s at 93°C, 10 s at 52°C, 60 s at 72°C) in 10 mM Tris-Cl (pH 8.4) containing 50 mM KCl, 7.5 mM MgCl₂, 0.05 mg/ml bovine serum albumin, 1 mM deoxynucleoside triphosphates, 5 μM primers (Table 1) (SEQ ID NOS: 2, 3) and 0.1 units/μl Taq polymerase. The 5' PCR primer was 5' end-labeled with polynucleotide kinase and [α-³²P]ATP and the 3' PCR primer was biotinylated at the 5' end using biotin phosphoramidite (Glen Research, Sterling, VA). Following PCR amplification, streptavidin (Pierce, Rockford, IL) was added to the unpurified PCR reaction mixture at a 10-fold molar excess over the biotinylated primer and incubated for 15 min at room temperature. The dsDNA was denatured by adding an equal volume of stop solution (90% formamide, 1% sodium dodecyl sulfate, 0.025% bromophenol blue and xylene cyanol) and incubating for 20 min at room temperature. The radiolabeled strand was separated from the streptavidin-bound biotinylated strand by electrophoresis on 12% polyacrylamide/7M urea gels. The faster migrating radiolabeled (non-biotinylated) ssDNA strand was cut out of the gel and recovered as described above. The amount of ssDNA was

estimated from the absorbance at 260 nm using the extinction coefficient of 33 $\mu\text{g/ml/absorbance unit}$ (Sambrook *et al.* (1989) Molecular Cloning: A Laboratory Manual, 2nd Ed. 3 vols., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York).

- 5 Cloning and Sequencing. The amplified affinity-enriched pool from SELEX round 12 was purified on a 12% polyacrylamide gel and cloned between *Hind*III and *Pst*I sites in JM109 strain of *E. coli* (Sambrook *et al.* (1989) Molecular Cloning: A Laboratory Manual, 2nd Ed. 3 vols., Cold Spring Harbor Laboratory Press, Cold Spring Harbor). Individual clones were used to prepare plasmids by alkaline lysis. Plasmids were sequenced at the insert region
10 using the forward sequencing primer and Sequenase 2.0 (Amersham, Arlington Heights, IL) according to the manufacturer's protocol.

- Determination of the apparent equilibrium dissociation constants and the dissociation rate constants. The binding of ssDNA ligands at low concentrations to varying concentrations of
15 PDGF was determined by the nitrocellulose filter binding method as described (Green *et al.* (1995) *Chemistry and Biology* 2:683-695). The concentrations of PDGF stock solutions (in PBS) were determined from the absorbance readings at 280 nm using the following ϵ_{280} values calculated from the amino acid sequences (Gill and von Hippel (1989) *Anal. Biochem.* 182:319-326): 19,500 $\text{M}^{-1}\text{cm}^{-1}$ for PDGF-AA, 15,700 $\text{M}^{-1}\text{cm}^{-1}$ for PDGF-AB and 11,800
20 $\text{M}^{-1}\text{cm}^{-1}$ for PDGF-BB. ssDNA for all binding experiments were purified by electrophoresis on 8% (>80 nucleotides) or 12% (<40 nucleotides) polyacrylamide/7 M urea gels. All ssDNA ligands were heated at 90°C in binding buffer at high dilution (~1 nM) for 2 min and cooled on ice prior to further dilution into the protein solution. The binding mixtures were typically incubated for 15 min at 37°C before partitioning on nitrocellulose filters.

- 25 The binding of DNA ligands (L) to PDGF-AA (P) is adequately described with the bimolecular binding model for which the fraction of bound DNA at equilibrium (q) is given by eq. 1,

$$q = \frac{f/2[L]_t}{\{[P]_t + [L]_t + K_d - ([P]_t + [L]_t + K_d)^2 - 4[P]_t[L]_t\}^{1/2}} \quad (1)$$

where $[P]_t$ and $[R]_t$ are total protein and total DNA concentrations, K_d is the equilibrium dissociation constant and f is the efficiency of retention of protein-DNA complexes on nitrocellulose filters (Irvine *et al.* (1991) J. Mol. Biol. 222:739-761; Jellinek *et al.* (1993) Proc. Natl. Acad. Sci. USA 90:11227-11231).

The binding of DNA ligands to PDGF-AB and PDGF-BB is biphasic and can be described by a model in which the DNA ligand is composed of two non-interconverting components (L_1 and L_2) that bind to the protein with different affinities, described by corresponding dissociation constants, K_{d1} and K_{d2} (Jellinek *et al.* (1993) Proc. Natl. Acad. Sci. USA 90:11227-11231). In this case, the explicit solution for the fraction of bound DNA (q) is given by eq. 2,

$$q = f \left(\frac{X_1 K_{d1}}{1 + K_{d1} [P]} + \frac{X_2 K_{d2}}{1 + K_{d2} [P]} \right) [P] \quad (2)$$

with

$$[P] = \frac{[P]_t}{1 + \frac{X_1 K_{d1} [L]_t}{1 + K_{d1} [P]} + \frac{X_2 K_{d2} [L]_t}{1 + K_{d2} [P]}}$$

where X_1 and $X_2(=1-X_1)$ are the mole fractions of L_1 and L_2 . The K_d values for the binding of DNA ligands to PDGF were calculated by fitting the data points to eq. 1 (for PDGF-AA) or eq. 2 (for PDGF-AB and PDGF-BB) using the non-linear least squares method.

The dissociation rate constants (k_{off}) were determined by measuring the amount of ^{32}P 5'-end labeled minimal ligands (0.17 nM) bound to PDGF-AB (1 nM) as a function of time following the addition of 500-fold excess of unlabeled ligands, using nitrocellulose filter binding as the partitioning method. The k_{off} values were determined by fitting the data points to the first-order rate equation (eq. 3)

$$(q-q_8)/(q_0-q_8) = \exp(-k_{\text{off}}t) \quad (3)$$

where q , q_0 and q_8 represent the fractions of DNA bound to PDGF-AB at any time (t), $t=0$
 5 and $t=8$, respectively.

Minimal ligand determinations. To generate a population of 5' end-labeled DNA ligands serially truncated from the 3' end, a primer complementary to the 3' invariant sequence region of a DNA ligand template (truncated primer 5N2, Table 1) (SEQ ID NO: 3) was radiolabeled
 10 at the 5' end with $[\gamma\text{-}^{32}\text{P}]\text{-ATP}$ and T4 polynucleotide kinase, annealed to the template and extended with Sequenase (Amersham, Arlington Heights, IL) and a mixture of all four dNTPs and ddNTPs. Following incubation in binding buffer for 15 min at 37°C, the fragments from this population that retain high affinity binding to PDGF-AB were separated from those with weaker affinity by nitrocellulose filter partitioning. Electrophoretic
 15 resolution of the fragments on 8% polyacrylamide/7 M urea gels, before and after affinity selection, allows determination of the 3' boundary. To generate a population of 3' end-labeled DNA ligands serially truncated from the 5' end, the DNA ligands were radiolabeled at the 3' end with $[\alpha\text{-}^{32}\text{P}]\text{-cordycepin-5'-triphosphate}$ (New England Nuclear, Boston, MA) and T4 RNA ligase (Promega, Madison, WI), phosphorylated at the 5' end with
 20 ATP and T4 polynucleotide kinase, and partially digested with lambda exonuclease (Gibco BRL, Gaithersburg, MD). Partial digestion of 10 pmols of 3'-labeled ligand was done in 100 μL volume with 7 mM glycine-KOH (pH 9.4), 2.5mM MgCl_2 , 1 $\mu\text{g/ml}$ BSA, 15 μg tRNA, and 4 units of lambda exonuclease for 15 min at 37°. The 5' boundary was determined in an analogous manner to that described for the 3' boundary.

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Melting temperature (T_m) measurements. Melting profiles for the minimal DNA ligands were obtained on a Cary Model 1E spectrophotometer. Oligonucleotides (320-400 nM) were heated to 95°C in PBS, PBSM or PBS with 1 mM EDTA and cooled to room temperature prior to the melting profile determination. Melting profiles were generated by heating the
 30 samples at the rate of 1 °C/min from 15-95°C and recording the absorbance every 0.1 °C. The

first derivative of the data points was calculated using the plotting program KaleidaGraph (Synergy Software, Reading, PA). The first derivative values were smoothed using a 55 point smoothing function by averaging each point with 27 data points on each side. The peak of the smoothed first derivative curves was used to estimate the T_m values.

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Crosslinking of 5-iodo-2'-deoxyuridine-substituted DNA ligands to PDGF-AB. DNA ligands containing single or multiple substitutions of 5'-iodo-2'-deoxyuridine for thymidine were synthesized using the solid phase phosphoramidite method. To test for the ability to crosslink, trace amounts of 5' ^{32}P end-labeled ligands were incubated with PDGF-AB (100 nM) in binding buffer at 37° C for 15 min prior to irradiation. The binding mixture was transferred to a 1 cm path length cuvette thermostated at 37° C and irradiated at 308 nm for 25-400 s at 20 Hz using a XeCl charged Lumonics Model EX748 excimer laser. The cuvette was positioned 24 cm beyond the focal point of a convergent lens, with the energy at the focal point measuring 175 mjoules/pulse. Following irradiation, aliquots were mixed with an equal volume of formamide loading buffer containing 0.1% SDS and incubated at 95° C for 5 min prior to resolution of the crosslinked PDGF/ligand complex from the free ligand on 8% polyacrylamide/7 M urea gels.

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To identify the protein site of crosslinking for ligand 20t-I4 (SEQ ID NO: 92), binding and irradiation were done on a larger scale. PDGF-AB and 5' ^{32}P end-labeled ligand, each at 1 μM in PBSM, were incubated and irradiated (300 s) as described above in two 1 ml reaction vessels. The reaction mixtures were combined, ethanol precipitated and resuspended in 0.3 ml of Tris-HCl buffer (100 mM, pH 8.5). The PDGF-AB/ligand crosslinked complex was digested with 0.17 $\mu\text{g}/\mu\text{l}$ of modified trypsin (Boehringer Mannheim) for 20 hours at 37°C. The digest mixture was extracted with phenol/chloroform, chloroform and then ethanol precipitated. The pellet was resuspended in water and an equal volume of formamide loading buffer with 5% (v/v) β -mercaptoethanol (no SDS), incubated at 95° C for 5 min, and resolved on a 40 cm 8% polyacrylamide/7 M urea gel. The crosslinked tryptic-peptide/ligand that migrated as two closely spaced bands about 1.5 cm above the free ligand band was excised from the gel and eluted by the crush and soak method and ethanol precipitated. The

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dried crosslinked peptide (about 160 pmoles based on the specific activity) was sequenced by Edman degradation (Midwest Analytical, Inc., St. Louis, MO).

Receptor Binding Assay. The binding of ^{125}I -PDGF-AA and ^{125}I -PDGF-BB to porcine aortic endothelial (PAE) cells transfected with PDGF α - or β -receptors were performed as described (Heldin *et al.*, (1988) EMBO J. 7:1387-1394). Different concentrations of DNA ligands were added to the cell culture (1.5 cm^2) in 0.2 ml of phosphate buffered saline supplemented with 1 mg bovine serum albumin per ml together with ^{125}I -PDGF-AA (2 ng, 100,000 cpm) or ^{125}I -PDGF-BB (2 ng, 100,000 cpm). After incubation at 4°C for 90 min, the cell cultures were washed and cell associated radioactivity determined in a g-counter (Heldin *et al.*, (1988) EMBO J. 7:1387-1394).

^3H thymidine Incorporation Assay. The incorporation of ^3H thymidine into PAE cells expressing PDGF β -receptor in response to 20 ng/ml of PDGF-BB or 10% fetal calf serum and in the presence of different concentrations of DNA ligands was performed as described (Mori *et al.* (1991) J. Biol. Chem. 266:21158-21164). After incubation for 24 h at 37°C , ^3H -radioactivity incorporated into DNA was determined using a β -counter.

EXAMPLE 2. ssDNA LIGANDS OF PDGF

High affinity DNA ligands to PDGF AB were identified by the SELEX process from a library of $\sim 3 \times 10^{14}$ molecules (500 pmol) of single stranded DNA randomized at forty contiguous positions (Table 1) (SEQ ID NO: 1). The PDGF-bound DNA was separated from unbound DNA by polyacrylamide gel electrophoresis in the first round and by nitrocellulose filter binding in the subsequent rounds. After 12 rounds of SELEX, the affinity-enriched pool bound to PDGF-AB with an apparent dissociation constant (K_d) of $\sim 50\text{ pM}$ (data not shown). This represented an improvement in affinity of ~ 700 -fold compared to the initial randomized DNA library. This affinity-enriched pool was used to generate a cloning library from which 39 isolates were sequenced. Thirty-two of these ligands were found to have unique sequences (Table 2) (SEQ ID NOS: 4-35). Ligands that were subjected to the minimal sequence determination are marked with an asterisk (*) next to the clone number.

The clone numbers that were found to retain high affinity binding as minimal ligands are italicized. All ligands shown in Table 2 were screened for their ability to bind to PDGF AB using the nitrocellulose filter binding method. To identify the best ligands from this group, the relative affinities for PDGF-AB were determined by measuring the fraction of 5' ³²P end-labeled ligands bound to PDGF-AB over a range of protein concentrations. For the ligands that bound to PDGF-AB with high affinity, the affinity toward PDGF-BB and PDGF-AA was also examined: in all cases, the affinity of ligands for PDGF-AB and PDGF-BB was comparable while the affinity for PDGF-AA was considerably lower (data not shown).

Twenty-one of the thirty-two unique ligands can be grouped into a sequence family shown in Table 3 (SEQ ID NOS: 4, 5, 7-9, 14-24, 26, 31, 32, 34 and 35). The sequences of the initially randomized region (uppercase letters) are aligned according to the consensus three-way helix junction motif. Nucleotides in the sequence-invariant region (lowercase letters) are only shown where they participate in the predicted secondary structure. Several ligands were "disconnected" (equality symbol) in order to show their relatedness to the consensus motif through circular permutation. The nucleotides predicted to participate in base pairing are indicated with underline inverted arrows, with the arrow heads pointing toward the helix junction. The sequences are divided into two groups, A and B, based on the first single stranded nucleotide (from the 5' end) at the helix junction (A or G, between helices II and III). Mismatches in the helical regions are shown with dots under the corresponding letters (G-T and T-G base pairs were allowed). In places where single nucleotide bulges occur, the mismatched nucleotide is shown above the rest of the sequence between its neighbors.

This classification is based in part on sequence homology among these ligands, but in greater part on the basis of a shared secondary structure motif: a three-way helix junction with a three nucleotide loop at the branch point (Figure 1) (SEQ ID NO: 82). These ligands were subdivided into two groups; for ligands in group A, the loop at the branch point has an invariant sequence AGC and in group B, that sequence is G(T/G)(C/T). The proposed consensus secondary structure motif is supported by base-pairing covariation at non-conserved nucleotides in the helices (Table 4). Since the three-way junctions are

encoded in continuous DNA strands, two of the helices end in loops at the distal end from the junction. These loops are highly variable, both in length and in sequence. Furthermore, through circular permutation of the consensus motif, the loops occur in all three helices, although they are most frequent in helices II and III. Together these observations suggest that the regions distal from the helix junction are not important for high affinity binding to PDGF-AB. The highly conserved nucleotides are indeed found near the helix junction (Table 3, Figure 1).

EXAMPLE 3. MINIMAL LIGAND DETERMINATIONS

The minimal sequence necessary for high affinity binding was determined for six of the best ligands to PDGF-AB. In general, the information about the 3' and 5' minimal sequence boundaries can be obtained by partially fragmenting the nucleic acid ligand and then selecting for the fragments that retain high affinity for the target. With RNA ligands, the fragments can be conveniently generated by mild alkaline hydrolysis (Tuerk *et al.* (1990) J. Mol. Biol. 213: 749-761; Jellinek *et al.* (1994) Biochemistry 33:10450-10456; Jellinek *et al.* (1995) Biochemistry 34:11363-11372; Green *et al.* (1995) J. Mol. Biol. 247:60-68). Since DNA is more resistant to base, an alternative method of generating fragments is needed for DNA. To determine the 3' boundary, a population of ligand fragments serially truncated at the 3' end was generated by extending the 5' end-labeled primer annealed to the 3' invariant sequence of a DNA ligand using the dideoxy sequencing method. This population was affinity-selected by nitrocellulose filtration and the shortest fragments (truncated from the 3' end) that retain high affinity binding for PDGF-AB were identified by polyacrylamide gel electrophoresis. The 5' boundary was determined in an analogous manner except that a population of 3' end-labeled ligand fragments serially truncated at the 5' end was generated by limited digestion with lambda exonuclease. The minimal ligand is then defined as the sequence between the two boundaries. It is important to keep in mind that, while the information derived from these experiments is useful, the suggested boundaries are by no means absolute since the boundaries are examined one terminus at the time. The untruncated (radiolabeled) termini can augment, reduce or have no effect on binding (Jellinek *et al.* (1994) Biochemistry 33:10450-10456).

Of the six minimal ligands for which the boundaries were determined experimentally, two (20t (SEQ ID NO: 83) and 41t (SEQ ID NO: 85); truncated versions of ligands 20 and 41) bound with affinities comparable (within a factor of 2) to their full-length analogs and four had considerably lower affinities. The two minimal ligands that retained high affinity binding to PDGF, 20t and 41t, contain the predicted three-way helix junction secondary structure motif (Figure 2) (SEQ ID NOS: 83-85). The sequence of the third minimal ligand that binds to PDGF-AB with high affinity, 36t, was deduced from the knowledge of the consensus motif (Figure 2). In subsequent experiments, we found that the single-stranded region at the 5' end of ligand 20t is not important for high affinity binding. Furthermore, the trinucleotide loops on helices II and III in ligand 36t (GCA and CCA) can be replaced with hexaethylene glycol spacers (*infra*). These experiments provide further support for the importance of the helix junction region in high affinity binding to PDGF-AB.

Binding of the Minimal Ligands to PDGF. The binding of minimal ligands 20t, 36t, and 41t to varying concentrations of PDGF-AA, PDGF-AB and PDGF-BB is shown in Figures 3A-3C. In agreement with the binding properties of their full length analogs, the minimal ligands bind to PDGF-AB and PDGF-BB with substantially higher affinity than to PDGF AA (Figures 3A-3C, Table 5). In fact, their affinity for PDGF-AA is comparable to that of random DNA (data not shown). The binding to PDGF-AA is adequately described with a monophasic binding equation while the binding to PDGF-AB and PDGF-BB is notably biphasic. In previous SELEX experiments, biphasic binding has been found to be a consequence of the existence of separable nucleic acid species that bind to their target protein with different affinities (Jellinek *et al.* (1995) *Biochemistry* 34:11363-11372) and unpublished results). The identity of the high and the low affinity fractions is at present not known. Since these DNA ligands described here were synthesized chemically, it is possible that the fraction that binds to PDGF-AB and PDGF-BB with lower affinity represents chemically imperfect DNA. Alternatively, the high and the low affinity species may represent stable conformational isomers that bind to the PDGF B-chain with different affinities. In any event, the higher affinity binding component is the most populated ligand species in all cases (Figures 3A-3C). For comparison, a 39-mer DNA ligand that binds to

human thrombin with a K_d of 0.5 nM (ligand T39 (SEQ ID NO.: 88)):
5'-CAGTCCGTGGTAGGGCAGGTTGGGGTGA CTTTCGTGGAA[3'T], where [3'T]
represents a 3'-3' linked thymidine nucleotide added to reduce 3'-exonuclease degradation
and has a predicted stem-loop structure, binds to PDGF-AB with a K_d of 0.23 μ M (data not
5 shown).

Dissociation Rates of the Minimal Ligands. To evaluate the kinetic stability of the
PDGF-AB/DNA complexes, the dissociation rates at 37°C for the complexes of minimal
ligands 20t, 36t and 41t with PDGF-AB were determined by measuring the amount of
10 radiolabeled ligands (0.17 nM) bound to PDGF-AB (1 nM) as a function of time following
the addition of a large excess of unlabeled ligands (Figure 4). At these protein and DNA
ligand concentrations, only the high affinity fraction of the DNA ligands binds to PDGF-AB.
The following values for the dissociation rate constants were obtained by fitting the data
points shown in Figure 4 to the first-order rate equation: $4.5 \pm 0.2 \times 10^{-3} \text{ s}^{-1}$ ($t_{1/2} = 2.6 \text{ min}$)
15 for ligand 20t, $3.0 \pm 0.2 \times 10^{-3} \text{ s}^{-1}$ ($t_{1/2} = 3.8 \text{ min}$) for ligand 36t, and $1.7 \pm 0.1 \times 10^{-3} \text{ s}^{-1}$ ($t_{1/2} =$
6.7 min) for ligand 41t. The association rates calculated for the dissociation constants and
dissociation rate constants ($k_{on}=k_{off}/K_d$) are $3.1 \times 10^7 \text{ M}^{-1}\text{s}^{-1}$ for 20t, $3.1 \times 10^7 \text{ M}^{-1}\text{s}^{-1}$ for 36t
and $1.2 \times 10^7 \text{ M}^{-1}\text{s}^{-1}$ for 41t.

Melting Temperatures of the Minimal Ligands. Melting temperatures (T_m 's) were determined
for minimal ligands 20t, 36t and 41t from the UV absorption vs. temperature profiles (Figure
5). At the oligonucleotide concentrations used in these experiments (320-440 nM), only the
monomeric species were observed as single bands on non-denaturing polyacrylamide gels.
The T_m values were obtained from the first derivative replots of the melting profiles. Ligands
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25 20t and 41t exhibited monophasic melting with T_m values of 44 °C and 49 °C. The melting
profile of ligand 36t was biphasic, with the T_m value of 44 °C for the first (major) transition
and ~63 °C for the second transition.

Photocrosslinking of 5-Iodo-2'-Deoxyuridine Substituted Minimal DNA Ligands to PDGF-
30 AB. To determine the sites on the DNA ligands and PDGF that are in close contact, a series

of photo-crosslinking experiments was performed with 5'-iodo-2'-deoxyuridine (IdU)-substituted DNA ligands 20t, 36t and 41t. Upon monochromatic excitation at 308 nm, 5-iodo- and 5-bromo-substituted pyrimidine nucleotides populate a reactive triplet state following intersystem crossing from the initial n to π^* transition. The excited triplet state species then reacts with electron rich amino acid residues (such as Trp, Tyr and His) that are in its close proximity to yield a covalent crosslink. This method has been used extensively in studies of nucleic acid-protein interactions since it allows irradiation with >300 nm light which minimizes photodamage (Willis *et al.* (1994) *Nucleic Acids Res.* 22:4947-4952; Stump and Hall (1995) *RNA* 1:55-63; Willis *et al.* (1993) *Science* 262:1255-1257; Jensen *et al.* (1995) *Proc. Natl. Acad. Sci. U. S. A.* 92:12220-12224). Analogs of ligands 20t, 36t and 41t were synthesized in which all thymidine residues were replaced with IdU residues using the solid phase phosphoramidite method. The affinity of these IdU-substituted ligands for PDGF-AB was somewhat enhanced compared to the unsubstituted ligands and based on the appearance of bands with slower electrophoretic mobility on 8% polyacrylamide/7 M urea gels, all three 5' end-labeled IdU-substituted ligands crosslinked to PDGF-AB upon irradiation at 308 nm (data not shown). The highest crosslinking efficiency was observed with IdU-substituted ligand 20t. To identify the specific IdU position(s) responsible for the observed crosslinking, seven singly or multiply IdU-substituted analogs of 20t were tested for their ability to photo-crosslink to PDGF-AB: ligands 20t-I1 through 20t-I7 (5'-TGGGAGGGCGCGT¹T¹CT¹T¹CGT²GGT³T⁴ACT⁵T⁶T⁶T⁶AGT⁷CCCG-3' (SEQ ID NOS.: 89-95) where the numbers indicate IdU substitutions at indicated thymidine nucleotides for the seven ligands). Of these seven ligands, efficient crosslinking to PDGF-AB was observed only with ligand 20t-I4 (SEQ ID NO: 92). The photo-reactive IdU position corresponds to the 3' proximal thymidine in the loop at the helix junction (Figure 2).

To identify the crosslinked amino acid residue(s) on PDGF-AB, a mixture of 5' end-labeled 20t-I4 and PDGF-AB was incubated for 15 min at 37°C followed by irradiation at 308 nm. The reaction mixture was then digested with modified trypsin and the crosslinked fragments resolved on an 8% polyacrylamide/7 M urea gel. Edman degradation of the peptide fragment recovered from the band that migrated closest to the free DNA band revealed the amino acid sequence KKPIXKK (SEQ ID NO: 96), where X indicates a

modified amino acid that could not be identified with the 20 derivatized amino acid standards. This peptide sequence, where X is phenylalanine, corresponds to amino acids 80-86 in the PDGF-B chain (Johnsson *et al.* (1984) EMBO J. 3:921-928) which in the crystal structure of PDGF-BB comprises a part of solvent-exposed loop III (Oefner *et al.* (1992) EMBO J. 11:3921-3926). In the PDGF A-chain, this peptide sequence does not occur (Betsholtz *et al.* (1986) Nature 320:695-699). Together, these data establish a point contact between a specific thymidine residue in ligand 20t and phenylalanine 84 of the PDGF B-chain.

Receptor Binding Assay. In order to determine whether the DNA ligands to PDGF were able to inhibit the effects of PDGF isoforms on cultured cells, the effects on binding of ¹²⁵I-labeled PDGF isoforms to PDGF α - and β -receptors stably expressed in porcine aortic endothelial (PAE) cells by transfection were first determined. Ligands 20t, 36t and 41t all efficiently inhibited the binding of ¹²⁵I-PDGF-BB to PDGF α -receptors (Figure 6) or PDGF β -receptors (data not shown), with half maximal effects around 1 nM of DNA ligand. DNA ligand T39 (described *supra*), directed against thrombin and included as a control, showed no effect. None of the ligands was able to inhibit the binding of ¹²⁵I-PDGF-AA to the PDGF α -receptor (data not shown), consistent with the observed specificity of ligands 20t, 36t and 41t for PDGF-BB and PDGF-AB.

Inhibition of Mitogenic Effects by Minimal Ligands. The ability of the DNA ligands to inhibit the mitogenic effects of PDGF-BB on PAE cells expressing PDGF β -receptors was investigated. As shown in Figure 7, the stimulatory effect of PDGF-BB on [³H]thymidine incorporation was neutralized by ligands 20t, 36t and 41t. Ligand 36t exhibited half maximal inhibition at the concentration of 2.5 nM; ligands 41t was slightly more efficient and 20t slightly less efficient. The control ligand T39 had no effect. Moreover, none of the ligands inhibited the stimulatory effects of fetal calf serum on [³H]thymidine incorporation in these cells, showing that the inhibitory effects are specific for PDGF.

EXAMPLE 4. POST-SELEX MODIFICATIONS

The stability of nucleic acids to nucleases is an important consideration in efforts to develop nucleic acid-based therapeutics. Experiments have shown that many, and in some cases most of the nucleotides in SELEX-derived ligands can be substituted with modified nucleotides that resist nuclease digestion, without compromising high affinity binding (Green *et al.* (1995) *Chemistry and Biology* 2:683-695; Green *et al.* (1995) *J. Mol. Biol.* 247:60-68).

A series of substitution experiments were conducted to identify positions in ligand 36t that tolerate 2'-O-methyl (2'-O-Me) or 2'-fluoro (2'-F) substitution. Tables 6 and 7 and Figures 8A and 8B summarize the substitutions examined and their effect on the affinity of the modified ligands for PDGF-AB or PDGF-BB. 2-Fluoropyrimidine nucleoside phosphoramidites were obtained from JBL Scientific (San Louis Obispo, CA). 2'-O-Methylpurine phosphoramidites were obtained from PerSeptive Biosystems (Boston, MA). All other nucleoside phosphoramidites were from PerSeptive Biosystems (Boston, MA). Not all substitution combinations were examined. Nevertheless, these experiments have been used to identify the pattern of 2'-O-Me and 2'-F substitutions that are compatible with high affinity binding to PDGF-AB or PDGF-BB. It is worth noting that trinucleotide loops on helices II and III in ligand 36t (Figures 2 and 8B) can be replaced with pentaethylene glycol (18-atom) spacers (Spacer Phosphoramidite 18, Glen Research, Sterling, VA) (see Example 5 for description of synthesis of pentaethylene glycol-substituted ligand) without compromising high affinity binding to PDGF-AB or -BB. This is in agreement with the notion that the helix junction domain of the ligand represents the core of the structural motif required for high affinity binding. In practical terms, the replacement of six nucleotides with two pentaethylene glycol spacers is advantageous in that it reduces by four the number of coupling steps required for the synthesis of the ligand. In addition to the substitution experiments, four nucleotides from the base of helix I were found that could be deleted without loss of binding affinity (compare for example ligand 36t with 36ta or ligand 1266 with 1295 in Tables 6 and 7).

EXAMPLE 5. SYNTHESIS OF PEG-MODIFIED PDGF NUCLEIC ACID LIGANDS

A) General Procedure for the Synthesis of NX31975 on Solid Support

Synthesis was carried out on 1mmol scale on a millipore 8800 automated synthesizer using standard deoxynucleoside phosphoramidites, 2'-O-methyl-5'-O-DMT-N2-tert-butylphenoxyacetylguanosine-phosphoramidite, 2'-O-methyl-5'-O-DMT-N6-tert-butylphenoxyacetyl-adenosine-phosphoramidite, 2'-deoxy-2'-fluoro-5'-O-DMT-uridine-phosphoramidite, 2'-deoxy-2'-fluoro-5'-O-DMT-N4-acetylcytidine-3'-N,N-diisopropyl-(2-cyanoethyl)-phosphoramidite, 18-O-DMT-hexaethyleneglycol-1-[N,N-diisopropyl-(2-cyanoethyl)-phosphoramidite,] (Figure 9C), and 5-trifloroacetamidopentane-1-[N,N-diisopropyl-(2-cyanoethyl)-phosphoramidite,]. (Figure 9D). The syntheses were carried out using 4,5-dicyanoimidazole as the activator on controlled pore glass (CPG) support of 600A pore size, 80-120 mesh, and 60 -70 $\mu\text{mol/g}$ loading with 5'-succinyl thymidine.

After the synthesis, the oligos were deprotected with 40% NH_4OH , at 55 $^{\circ}\text{C}$ for 16h. The support was filtered, and washed with water and 1:1 acetonitrile/water and the combined washings were evaporated to dryness. The ammonium counterion on the backbone was exchanged for triethylammonium ion by reverse phase salt exchange and the solvent was evaporated to afford the crude oligo as the triethylammonium salt.

Hexaethylene glycol spacers on the loops are attached to the nucleotides through phosphate linkages. The structures of the 2 loops are shown in Figures 9A and 9B. The 5' phosphate group shown is from the hexaethylene glycol phosphoramidite.

B) Conjugation of 40K PEG NHS ester to the aminolinker on PDGF Nucleic Acid Ligands

The NX31975 crude oligonucleotide containing the 5' primary amino group was dissolved in 100mM sodium borate buffer (pH 9) to 60 mg /ml concentration. In a separate tube 2 Eq of PEG NHS ester (Figure 9E) (Shearwater Polymers, Inc.) was dissolved in dry DMF (Ratio of borate : DMF 1:1) and the mixture was warmed to dissolve the PEG NHS ester. Then the oligo solution was quickly added to PEG solution and the mixture was vigorously stirred at room temperature for 10 minutes. About 95% of the oligo conjugated to the PEG NHS ester.

EXAMPLE 6. STABILITY OF MODIFIED LIGANDS IN SERUM

The stabilities of DNA (36ta) and modified DNA (NX21568) ligands in rat serum at 37°C were compared. Serum used for these experiments was obtained from a Sprague-Dawley rat and was filtered through 0.45 µm cellulose acetate filter and buffered with 20 mM sodium phosphate buffer. Test ligands (36ta or NX21568) were added to the serum at the final concentration of 500 nM. The final serum concentration was 85% as a result of the addition of buffer and ligand. From the original 900 µl incubation mixture, 100 µl aliquots were withdrawn at various time points and added to 10 µl of 500 mM EDTA (pH 8.0), vortexed and frozen on dry ice and stored at -20°C until the end of the experiment. The amount of full length oligonucleotide ligand remaining for each of the time points was quantitated by HPLC analysis. To prepare the samples for HPLC injections, 200 µl of a mixture of 30% formamide, 70% 25 mM Tris buffer (pH 8.0) containing 1% acetonitrile was added to 100 µl of thawed time point samples, vortexed for 5 seconds and centrifuged for 20 minutes at 14,000 rpm in an Eppendorf microcentrifuge. The analysis was performed using an anion exchange chromatography column (NuceoPac, Dionex, PA-100, 4x50 mm) applying a LiCl gradient. The amount of full length oligonucleotide remaining at each time point was determined from the peak areas (Figure 10). With a half-life of about 500 min, the modified ligand (NX21568) exhibited a substantially greater stability in rat serum compared with the DNA ligand (36ta), which was degraded with a half-life of about 35 min (Figure 10). Thus, the increase in stability in serum results from the 2'-substitutions.

EXAMPLE 7. EFFICACY OF NX31975-40K PEG IN RESTENOSIS

Rat Restenosis Model and Efficacy Results. The plasma residence time of Nucleic Acid Ligands is dramatically improved by the addition of large, inert functional groups such as polyethylene glycol (see for example PCT/US 97/18944). For *in vivo* efficacy experiments, 40K PEG was conjugated to NX31975 to create NX31975 40K PEG as described in Example 5B (see Figure 9A for molecular description). Importantly, based on binding experiments, the addition of 40 kDa PEG group at the 5'-end of the ligand does not affect its binding affinity for PDGF-BB.

The effect of selective inhibition of PDGF-B by NX31975-40K PEG was studied in three-month-old male Sprague-Dawley rats (370-450 g). The rats were housed three to a cage with free access to a standard laboratory diet and water. Artificial light was provided 14 hours per day. The experiments were performed in accordance with the institutional
5 guidelines at the Animal Department, Department of Surgery, University Hospital, Uppsala University, Sweden.

A total of 30 rats were randomly allocated to one of two treatment groups: 15 rats in group one received 10 mg/kg body weight of NX31975-40K PEG in phosphate buffered saline (PBS) twice daily delivered by intraperitoneal (i.p.) injections and 15 rats in group two
10 (the control group) received an equal volume of PBS (about 1 ml). The duration of treatment was 14 days. The first injections in both groups were given one hour before arterial injury.

To generate the arterial lesions, all animals were anaesthetized with an i.p. injection of a mixture of one part Fentanyl-fluanisone (Hypnorm vet, fluanisone 10 mg/ml, fentanyl
15 0.2 mg/ml, Janssen Pharmaceutica, Beerse, Belgium), one part midazolam (Dormicum, Midazolam 5 mg/ml. F. Hoffman-La Roche AG, Basel, Switzerland) and two parts sterile water, 0.33 ml/100 g rat. The distal left common carotid and external carotid arteries were exposed through a midline incision in the neck. The left common carotid artery was traumatized by intraluminal passage of 2F Fogarty embolectomy catheter introduced through
20 the external carotid artery. The catheter was passed three times with the balloon expanded sufficiently with 0.06 ml distilled water to achieve a distension of the carotid itself. The external carotid was ligated after removal of the catheter and the wound was closed. All surgical procedures were performed by a surgeon blinded to the treatment groups.

Fourteen days after the catheter injury, the animals were anesthetized as above.
25 Twenty minutes before the exposure of the abdominal aorta the animals received an intravenous injection of 0.5 ml 0.5% Evans blue dye (Sigma Chemical Co., St. Louis, MO) to allow identification of the vessel segment which remained deendothelialized. The carotid arteries were perfused with ice-chilled PBS in situ at 100 mm Hg, via a large cannula placed retrograde in the abdominal aorta until the effluent ran clear via inferior caval vein vent. A
30 distal half of the right and left common carotid arteries, up to the level of the bifurcation,

were removed and frozen in liquid nitrogen. Immediately thereafter, the remaining proximal segment was perfusion-fixed through the same aortic cannula at 100 mm Hg pressure with 2.5% glutaraldehyde in phosphate buffer, pH 7.3. Before starting perfusion with PBS, the animals were killed by an overdose of phenobarbital. After approximately 15 minutes of perfusion fixation, the remaining proximal right and left common carotid arteries were retrieved for further preparation, including the aortic arch and innominate artery.

Five sections, approximately 0.5 μ m apart, from the middle of the Evans blue stained segment of the left common carotid artery and one section from the contralateral uninjured artery were analyzed per animal with computer-assisted planimetry. The following areas were measured: the area encircled by external elastic lumina (EEL), internal elastic lumina (IEL) and the endoluminal cell layer. Areas for tunical media and tunica intima were calculated. All measurements by an individual blinded to the treatment regimens.

Based on values of intima/media ratios for the control and the Nucleic Acid Ligand-treated groups, the PDGF Nucleic Acid Ligand significantly ($p < 0.05$) inhibited about 50% of the neointima formation (Figure 11).

EXAMPLE 8. ANTAGONISM OF PDGF IN GLOMERULONEPHRITIS BY NX31975-40K PEG

This example provides the general procedures followed and incorporated in Example 9.

Materials and Methods

All Nucleic Acid Ligands and their sequence-scrambled controls were synthesized by the solid phase phosphoramidite method on controlled pore glass using an 8800 Milligen DNA Synthesizer and deprotected using ammonium hydroxide at 55°C for 16 h. The Nucleic Acid Ligand used in experiments described in this example and Example 9 is NX31975 40K PEG (Figure 9A). NX31975 40K PEG was created by conjugating NX31975 (Table 7) to 40K PEG as described in Example 5. In the sequence-scrambled control Nucleic Acid Ligand, eight nucleotides in the helix junction region of NX31975 were interchanged without

formally changing the consensus secondary structure (see Figure 8C). The binding affinity of the sequence-scrambled control Nucleic Acid Ligand for PDGF BB is $\approx 1\mu\text{M}$, which is 10,000 fold lower compared to NX21617. The sequence-scrambled control Nucleic Acid Ligand was then conjugated to PEG and named NX31976 40K PEG (see Figure 9B for molecular description). The covalent coupling of PEG to the Nucleic Acid Ligand (or to the sequence-scrambled control) was accomplished as described in Example 5.

Rat PDGF-BB for cross-reactivity binding experiments was derived from *E. coli* transfected with sCR-Script Amp SK(+) plasmid containing the rat PDGF-BB sequence. Rat PDGF-BB sequence was derived rat lung poly A+ RNA (Clontech, San Diego, CA) through RT-PCR using primers that amplify sequence encoding the mature form of PDGF-BB. Rat PDGF-BB protein expression and purification was performed at R&D Systems.

Mesangial Cell Culture Experiments

Human mesangial cells were established in culture, characterized and maintained as described previously (Radeke *et al.* (1994) J. Immunol. 153:1281-1292). To examine the antiproliferative effect of the ligands on the cultured mesangial cells, cells were seeded in 96-well plates (Nunc, Wiesbaden, Germany) and grown to subconfluency. They were then growth-arrested for 48 hours in MCDB 302 medium (Sigma, Deisenhofen, Germany). After 48 hours various stimuli together with either 50 or 10 $\mu\text{g/ml}$ Nucleic Acid Ligand NX31975 40K PEG or 50 or 10 $\mu\text{g/ml}$ sequence-scrambled Nucleic Acid Ligand (NX31976 40K PEG) were added: medium alone, 100 ng/ml human recombinant PDGF-AA, -AB or -BB (kindly provided by J. Hoppe, University of Würzburg, Germany), 100 ng/ml human recombinant epidermal growth factor (EGF; Calbiochem, Bad Soden, Germany) or 100 ng/ml recombinant human fibroblast growth factor-2 (kindly provided by Synergen, Boulder, Colorado). Following 72 hours of incubation, numbers of viable cells were determined using 2,3-bis[2-methoxy-4-nitro-5-sulfo-phenyl]-2H-tetrazolium-5-carboxanilide (XTT; Sigma) as described (Lonnemann *et al.* (1995) Kidney Int. 51:837-844).

Experimental Design

Anti-Thy 1.1 mesangial proliferative glomerulonephritis was induced in 33 male Wistar rats (Charles River, Sulzfeld, Germany) weighing 150-160g by injection of 1 mg/kg monoclonal anti-Thy 1.1 antibody (clone OX-7; European Collection of Animal Cell Cultures, Salisbury, England). Rats were treated with Nucleic Acid Ligands or PEG (see below) from day 3 to 8 after disease induction. Treatment consisted of twice daily i.v. bolus injections of the substances dissolved in 400 μ l PBS, pH 7.4. The treatment duration was chosen to treat rats from about one day after the onset to the peak of mesangial cell proliferation (Floege *et al.* (1993) *Kidney Int. Suppl.* 39:S47-54). Four groups of rats were studied: 1) nine rats, who received NX31975 40K PEG (i.e., a total of 4 mg of the PDGF-B ligand coupled to 15.7 mg 40K PEG); 2) ten rats, who received an equivalent amount of PEG-coupled, scrambled Nucleic Acid Ligand (NX31976 40K PEG); 3) eight rats, who received an equivalent amount (15.7 mg) of 40K PEG alone; 4) six rats, who received 400 μ l bolus injections of PBS alone. Renal biopsies for histological evaluation were obtained on days 6 and 9 after disease induction. Twenty-four hour urine collections were performed from days 5 to 6 and 8 to 9 after disease induction. The thymidine analogue 5-bromo-2'-deoxyuridine (BrdU; Sigma, Deisenhofen, Germany; 100 mg/kg body weight) was injected intraperitoneally at 4 hours prior to sacrifice on day 9.

Normal ranges of proteinuria and renal histological parameter (see below) were established in 10 non-manipulated Wistar rats of similar age.

Renal Morphology

Tissue for light microscopy and immunoperoxidase staining was fixed in methyl Carnoy's solution (Johnson *et al.* (1990) *Am. J. Pathol.* 136:369-374) and embedded in paraffin. Four μ m sections were stained with the periodic acid Schiff (PAS) reagent and

counterstained with hematoxylin. In the PAS stained sections the number of mitoses within 100 glomerular tufts was determined.

Immunoperoxidase Staining

5 Four mm sections of methyl Carnoy's fixed biopsy tissue were processed by an indirect immunoperoxidase technique as described (Johnson *et al.* (1990) *Am. J. Pathol.* 136:369-374). Primary antibodies were identical to those described previously (Burg *et al.* (1997) *Lab. Invest.* 76:505-516; Yoshimura *et al.* (1991) *Kidney Int.* 40:470-476) and included a murine monoclonal antibody (clone 1A4) to α -smooth muscle actin; a murine
10 monoclonal antibody (clone PGF-007) to PDGF B-chain; a murine monoclonal IgG antibody (clone ED1) to a cytoplasmic antigen present in monocytes, macrophages and dendritic cells; affinity purified polyclonal goat anti-human/bovine type IV collagen IgG preabsorbed with rat erythrocytes; an affinity purified IgG fraction of a polyclonal rabbit anti-rat fibronectin antibody; plus appropriate negative controls as described previously (Burg *et al.* (1997) *Lab.*
15 *Invest.* 76:505-516; Yoshimura *et al.* (1991) *Kidney Int.* 40:470-476). Evaluation of all slides was performed by an observer, who was unaware of the origin of the slides.

To obtain mean numbers of infiltrating leukocytes in glomeruli, more than 50 consecutive cross sections of glomeruli containing more than 20 discrete capillary segments were evaluated and mean values per kidney were calculated. For the evaluation of the
20 immunoperoxidase stains for α -smooth muscle actin, PDGF B-chain, type IV collagen and fibronectin each glomerular area was graded semiquantitatively, and the mean score per biopsy was calculated. Each score reflects mainly changes in the extent rather than intensity of staining and depends on the percentage of the glomerular tuft area showing focally enhanced positive staining: I = 0-25%, II = 25-50%, III = 50-75%, IV = >75%. This
25 semiquantitative scoring system is reproducible among different observers and the data are highly correlated with those obtained by computerized morphometry (Kliem *et al.* (1996) *Kidney Int.* 49:666-678; Hugo *et al.* (1996) *J. Clin. Invest.* 97:2499-2508).

Immunohistochemical Double-Staining

Double immunostaining for the identification of the type of proliferating cells was performed as reported previously (Kliem *et al.* (1996) *Kidney Int.* 49:666-678; Hugo *et al.* (1996) *J. Clin. Invest.* 97:2499-2508) by first staining the sections for proliferating cells with a murine monoclonal antibody (clone BU-1) against bromo-deoxyuridine containing nuclease in Tris buffered saline (Amersham, Braunschweig, Germany) using an indirect immunoperoxidase procedure. Sections were then incubated with the IgG₁ monoclonal antibodies 1A4 against α -smooth muscle actin and ED1 against monocytes/macrophages. Cells were identified as proliferating mesangial cells or monocytes/macrophages if they showed positive nuclear staining for BrdU and if the nucleus was completely surrounded by cytoplasm positive for α -smooth muscle actin. Negative controls included omission of either of the primary antibodies in which case no double-staining was noted.

***In situ* Hybridization for Type IV Collagen mRNA**

In situ hybridization was performed on 4 mm sections of biopsy tissue fixed in buffered 10% formalin utilizing a digoxigenin-labelled anti-sense RNA probe for type IV collagen (Eitner *et al.* (1997) *Kidney Int.* 51:69-78) as described (Yoshimura *et al.* (1991) *Kidney Int.* 40:470-476). Detection of the RNA probe was performed with an alkaline phosphatase coupled anti-digoxigenin antibody (Genius Nonradioactive Nucleic Acid Detection Kit, Boehringer-Mannheim, Mannheim, Germany) with subsequent color development. Controls consisted of hybridization with a sense probe to matched serial sections, by hybridization of the anti-sense probe to tissue sections which had been incubated with RNase A before hybridization, or by deletion of the probe, antibody or color solution described (Yoshimura *et al.* (1991) *Kidney Int.* 40:470-476). Glomerular mRNA expression was semiquantitatively assessed using the scoring system described above.

Miscellaneous Measurements

Urinary protein was measured using the Bio-Rad Protein Assay (Bio-Rad Laboratories GmbH, München, Germany) and bovine serum albumin (Sigma) as a standard.

5 Statistical Analysis

All values are expressed as means \pm SD. Statistical significance (defined as $p < 0.05$) was evaluated using ANOVA and Bonferroni t-tests.

EXAMPLE 9.

10 For all experiments reported here, the modified DNA Nucleic Acid Ligand was conjugated to 40K PEG as described in Examples 5 and 8 and shown in Figures 9A and 9B. Since most Nucleic Acid Ligands have molecular weights ranging between 8 to 12 kDa (the modified PDGF Nucleic Acid Ligand has MW of 10 kDa), the addition of a large inert molecular entity such as PEG dramatically improves the residence times of Nucleic Acid
15 Ligands *in vivo* (see for example PCT/US 97/18944). Importantly, the addition of the PEG moiety to the 5' end of the Nucleic Acid Ligand has no effect on the binding affinity of the Nucleic Acid Ligand for PDGF-BB ($K_d \approx 1 \times 10^{-10}$ M).

20 Cross-reactivity of Nucleic Acid Ligands for Rat PDGF-BB

The sequence of PDGF is highly conserved among species, and human and rat PDGF B-chain sequences are 89% identical (Herren et al. (1993) Biochim. Biophys. Acta 1173:294; Lindner et al. (1995) Circ. Res. 76:951). Nevertheless, in view of the high specificity of Nucleic Acid Ligands (Gold et al. (1995) Ann. Rev. Biochem. 64:763-797), the correct
25 interpretation of the *in vivo* experiments requires understanding of the binding properties of the Nucleic Acid Ligands to rat PDGF B-chain. We have therefore cloned and expressed the

mature form of rat PDGF-BB in E. coli. The PDGF Nucleic Acid Ligands bound to rat and human recombinant PDGF-BB with the same high affinity (data not shown).

PDGF B-Chain DNA-Ligand Specifically Inhibits Mesangial Cell Proliferation in vitro

5 In growth arrested mesangial cells, the effects of NX31975 40K PEG or the scrambled Nucleic Acid Ligand (NX31976 40K PEG) on growth factor induced proliferation were tested. Stimulated growth rates of the cells were not affected by the addition of scrambled Nucleic Acid Ligand (Figure 12). Fifty $\mu\text{g/ml}$ of NX31975 40K PEG significantly reduced PDGF-BB induced mesangial cell growth (Figure 12). PDGF-AB and -AA induced
10 mesangial cell growth also tended to be lower with NX31975 40K PEG, but these differences failed to reach statistical significance (Figure 12). In contrast, no effects of NX31975 40K PEG on either EGF or FGF-2 induced growth were noted. Similar effects were noted if the Nucleic Acid Ligands were used at a concentration of 10 $\mu\text{g/ml}$ (data not shown).

15 Effects of PDGF B-Chain DNA-Ligand in Rats with Anti-Thy 1.1 Nephritis

Following the injection of anti-Thy 1.1 antibody, PBS treated animals developed the typical course of the nephritis, which is characterized by early mesangiolysis and followed by a phase of mesangial cell proliferation and matrix accumulation on days 6 and 9 (Floege *et al.* (1993) Kidney Int. Suppl. 39:S47-54). No obvious adverse effects were noted following the
20 repeated injection of Nucleic Acid Ligands or PEG alone, and all rats survived and appeared normal until the end of the study.

In PAS stained renal sections the mesangioproliferative changes on days 6 and 9 after disease induction were severe and indistinguishable among rats receiving PBS, PEG alone or the scrambled Nucleic Acid Ligand (data not shown). Histological changes were markedly
25 reduced and almost normalized in the NX31975 40K PEG ligand treated group In order to (semi-)quantitatively evaluate the mesangioproliferative changes, various parameters were analyzed:

a) Reduction of Mesangial Cell Proliferation

Glomerular cell proliferation, as assessed by counting the number of glomerular mitoses, was not significantly different between the three control groups on days 6 and 9 (Figure 13A). As compared to rats receiving the scrambled Nucleic Acid Ligand, treatment with PDGF-B ligand led to a reduction of glomerular mitoses by 64% on day 6 and by 78% on day 9 (Figure 13A). To assess the treatment effects on mesangial cells, the renal sections for α -smooth muscle actin were immunostained, which is expressed by activated mesangial cells only (Johnson *et al.* (1991) J. Clin. Invest. 87:847-858). Again, there were no significant differences between the three control groups on days 6 and 9. However, the immunostaining scores of α -smooth muscle actin were significantly reduced on day 6 and 9 in the NX31975 40K PEG treated group (Figure 13D). To specifically determine whether mesangial cell proliferation was reduced, NX31975 40K PEG treated rats and scrambled Nucleic Acid Ligand treated rats were double immunostained for a cell proliferation marker (BrdU) and α -smooth muscle actin. The data confirmed a marked decrease of proliferating mesangial cells on day 9 after disease induction: 2.2 ± 0.8 BrdU-/ α -smooth muscle actin positive cells per glomerular cross section in PDGF-B aptamer treated rats versus 43.3 ± 12.4 cells in rats receiving the scrambled Nucleic Acid Ligand, i.e., a 95% reduction of mesangial cell proliferation. In contrast, no effect of the PDGF-B aptamer was noted on proliferating monocytes/macrophages on day 9 after disease induction (PDGF-B aptamer treated rats: 2.8 ± 1.1 BrdU+/ED-1+ cells per 100 glomerular cross sections; scrambled aptamer treated rats: 2.7 ± 1.8).

b) Reduced Expression of Endogenous PDGF B-Chain

By immunohistochemistry, the glomerular PDGF B-chain expression was markedly upregulated in all three control groups (Figure 13B), similar to previous observations (Yoshimura *et al.* (1991) Kidney Int. 40:470-476). In the NX31975 40K PEG treated group

the glomerular overexpression of PDGF B-chain was significantly reduced in parallel with the reduction of proliferating mesangial cells (Figure 13B).

c) Reduction of Glomerular Monocyte/Macrophage Influx

5 The glomerular monocyte/macrophage influx was significantly reduced in the NX31975 40K PEG treated rats as compared to rats receiving scrambled Nucleic Acid Ligand on days 6 and 9 after disease induction (Figure 13E).

d) Effects on Proteinuria

10 Moderate proteinuria of up to 147 mg/24 hrs was present on day 6 after disease induction in the 3 control groups (Figure 13C). Treatment with NX31975 40K PEG reduced the mean proteinuria on day 6, but this failed to reach statistical significance (Figure 13C). Proteinuria on day 9 after disease induction was low and similar in all four groups (Figure 13C).

e) Reduction of Glomerular Matrix Production and Accumulation

15 By immunohistochemistry, marked glomerular accumulation of type IV collagen and fibronectin was noted in all three control groups (Figure 14). The overexpression of both glomerular type IV collagen and fibronectin was significantly reduced NX31975 40K PEG
20 treated rats (Figure 14). In the latter, glomerular staining scores approached those observed in normal rats (Figure 14). By *in situ* hybridization, the decreased glomerular expression of type IV collagen in NX31975 40K PEG treated rats was shown to be associated with decreased glomerular synthesis of this collagen type (Figure 14).

**EXAMPLE 10. EXPERIMENTAL PROCEDURE FOR EVOLVING
2'-FLUORO-2'-DEOXYPYRIMIDINE RNA LIGANDS TO PDGF AND RNA
SEQUENCES OBTAINED.**

5 2'-FLUORO-2'-DEOXYPYRIMIDINE RNA SELEX

SELEX with 2'-fluoro-2'-deoxypyrimidine RNA targeting PDGF AB was done essentially as described previously (vide supra, and Jellinek *et al.*, 1993, 1994: supra) using the primer template set as shown in Table 8 (SEQ ID NOS: 36-38). Briefly, the 2'-fluoro-2'-
10 deoxypyrimidine RNA for affinity selections was prepared by in vitro transcription from synthetic DNA templates using T7 RNA polymerase (Milligan *et al.* (1987) Nucl. Acids Res. 15:8783). The conditions for in vitro transcription described in detail previously (Jellinek *et al.* (1994) supra) were used, except that higher concentration (3 mM) of the
15 2'-fluoro-2'-deoxypyrimidine nucleoside triphosphates (2'-F-UTP and 2'-F-CTP) was used compared to ATP and GTP (1 mM). Affinity selections were done by incubating PDGF AB with 2'-fluoro-2'-deoxypyrimidine RNA for at least 15 min at 37 °C in PBS containing 0.01% human serum albumin. Partitioning of free RNA from protein-bound RNA was done by nitrocellulose filtration as described (Jellinek *et al.*, 1993, 1994: supra). Reverse
20 transcription of the affinity-selected RNA and amplification by PCR were done as described previously (Jellinek *et al.* (1994) supra). Nineteen rounds of SELEX were performed, typically selecting between 1-12% of the input RNA. For the first eight rounds of selection, suramin (3-15 µM) was included in the selection buffer to increase the selection pressure. The affinity-enriched pool (round 19) was cloned and sequenced as described (Schneider *et al.* (1992) supra). Forty-six unique sequences have been identified, and the sequences are
25 shown in Table 9 (SEQ ID NOS: 39-81). The unique-sequence ligands were screened for their ability to bind PDGF AB with high affinity. While random 2'-fluoropyrimidine RNA (Table 8) bound to PDGF with a dissociation constant (K_d) of 35 ± 7 nM, many of the affinity-selected ligands bound to PDGF AB with ~100-fold higher affinities. Among the unique ligands, clones 9 ($K_d = 91 \pm 16$ pM), 11 ($K_d = 120 \pm 21$ pM), 16 ($K_d = 116 \pm 34$ pM),
30 23 ($K_d = 173 \pm 38$ pM), 25 ($K_d = 80 \pm 22$ pM), 37 ($K_d = 97 \pm 29$ pM), 38 ($K_d = 74 \pm 39$ pM), and 40 ($K_d = 91 \pm 32$ pM) exhibited the highest affinity for PDGF AB (binding of all of these

ligands to PDGF AB is biphasic and the K_d for the higher affinity binding component is given).

Table 1. Starting DNA and PCR primers for the ssDNA SELEX experiment.

		SEQ ID
		NO.
5	Starting ssDNA: 5'-ATCCGCCTGATTAGCGATACT[-40N-]ACTTGAGCAAAATCACCTGCAGGGG-3'	1
	PCR Primer 3N2*: 5'-BBBCCCCTGCAGGTGATTTTGCTCAAGT-3'	2
10	PCR Primer 5N2**: 5'-CCGAAGCTTAATACGACTCACTATAGGG <u>ATCCGCCTGATTAGCGATACT</u> -3'	3

*B=biotin phosphoramidite (e. g., Glen Research, Sterling, VA)

**For rounds 10, 11, and 12, the truncated PCR primer 5N2 (underlined) was used to amplify the template.

Table 2. Unique Sequences of the ssDNA high affinity ligands to PDGF.

5'-ATCCGCCTGATTAGCGATACT [40N]			SEQ ID	
5 ACTTGAGCAAAATCACCTGCAGGGG-3'			NO	
	*14	AGGCTTGACAAAGGGCACCATGGCTTAGTGGTCCTAGT		4
	*41	CAGGGCACTGCAAGCAATTGTGGTCCCAATGGGCTGAGT		5
10	6	CCAGGCAGTCATGGTCATTGTTTACAGTCGTGGAGTAGGT		6
	23	AGGTGATCCCTGCAAAGGCAGGATAACGTCCTGAGCATC		7
	2	ATGTGATCCCTGCAGAGGGAGGANACGTCTGAGCATC		8
	34	CACGTGATCCCATAAGGGCTGCGCAAAATAGCAGAGCATC		9
	8	GGTGGACTAGAGGGCAGCAAACGATCCTTGGTTAGCGTCC		10
15	1	GGTGCACGAGGGCTTACACAAACGTACACGTTTCCCCGC		11
	5	TGTCGGAGCAGGGGCGTACGAAAACCTTTACAGTTCCCCCG		12
	*40	AGTGGAACAGGGCACGGAGAGTCAAACCTTGGTTTCCCCC		13
	47	GTGGGTAGGGATCGGTGGATGCCTCGTCACTTCTAGTCCC		14
	18	GGGCGCCCTAAACAAAGGGTGGTCACTTCTAGTCCCAGGA		15
20	30	TCCGGGCTCGGGATTCGTGGTCACTTTCAGTCCCGGATATA		16
	*20	ATGGGAGGGCGCGTTCTTCGTGGTTACTTTTAGTCCCG		17
	35	ACGGGAGGGCACGTTCTTCGTGGTTACTTTTAGTCCCG		18
	13	GCTCGTAGGGGGCGATTCTTTCGCCGTTACTTCCAGTCCT		19
	16	GAGGCATGTTAACATGAGCATCGTCTCACGATCCTCAGCC		20
25	*36	CCACAGGCTACGGCACGTAGAGCATCACCATGATCCTGTG		21
	50	GCGGGCATGGCACATGAGCATCTCTGATCCCGCAATCCTC		22
	4	ACCGGGCTACTTCGTAGAGCATCTCTGATCCCGGTGCTCG		23
	44	AAAGGGCGAACGTAGGTCGAGGCATCCATTGGATCCCTTC		24
	24	ACGGGCTCTGTCACTGTGGCACTAGCAATAGTCCCGTCGC		25
30	7	GGGCAGACCTTCTGGACGAGCATCACCTATGTGATCCCG		26
	*26	AGAGGGGAAGTAGGCTGCCTGACTCGAGAGAGTCCTCCCG		27
	19	AGGGGTGCGAAACACATAATCCTCGCGGATTCCCATCGCT		28
	48	GGGGGGGCAATGGCGGTACCTCTGGTCCCCTAAATAC		29

Table 2 Cont.

	46	GCGGCTCAAAGTCCTGCTACCCGCAGCACATCTGTGGTC	30
	25	TTGGGCGTGAATGTCCACGGGTACCTCCGGTCCCAAAGAG	31
5	31	TCCGCGCAAGTCCCTGGTAAAGGGCAGCCCTAACTGGTC	32
	12	CAAGTTCCCCACAAGACTGGGGCTGTTCAAACCGCTAGTA	33
	15	CAAGTAGGGCGCGACACACGTCCGGGCACCTAAGGTCCCA	34
	*38	AAAGTCGTGCAGGGTCCCCTGGAAGCATCTCCGATCCCAG	35

10 * Indicates a boundary experiment was performed.

Italics indicate the clones that were found to retain high affinity binding as minimal ligands.

Table 3

HELIX I										
HELIX II			HELIX III							
SEQ ID NO: Group A										
8	2	=AGGG	AGGA	TACG	TCTG	AGC	ATCac3'	5'ATGTGAT	CCCTGCAG=	
23	4	ACCGGG	CTAC	TTC	GTAG	AGC	ATC	TCT	GAT	CCCGGTGCTCG
26	7	EGGG	CGACC	TTCT	GGACG	AGC	ATCAC	CTAT	GTGAT	CCCCG
20	16	CEGAGG	CATG	TTAA	CATG	AGC	ATCGT	CTC	ACGAT	CCTCAGCC
21	36	CCACAGG	CTACG	GCA	CGTAG	AGC	ATCA	CCA	TGAT	CCCTGTG
35	38	AAAGTCGTGCAGGG	TCC	CCT	GGA	AGC	ATC	TCC	GAT	CCCAGacLt
24	44	AAAGGG	CGAAC	GTA	GGTCG	AGGCATCC	ATT	GGAT	CCCTTC	
22	50	GCGGG	CATG	GCA	CATG	AGC	ATC	TCT	GAT	CCCGCAATCCTC
7	23	=AGG	CAGGATAAC	GTCCTG	AGC	ATCac3'	5'AGGTGAT	CCCTGCAA=		
9	34	=GGG	CTGC	GCAAAATA	GCAG	AGC	ATCac3'	5'CACGTGAT	CCCATAA=	
SEQ ID NO: Group B										
19	13	GCTCGTAGG	GGGCGA	TTCTT	TGGCC	GTT	ACT	TCC	AGT	CCTac
4	14	LactAGG	CTT	GACA	AAG	GGC	ACCAT	GGCTTAGTGGT	CCTAGTa	
34	15	CTCAAGTAGGG	CGGAC	ACAC	GTCCG	GGC	ACC	TAA	GGT	CCC ^a acttgag
15	18	CEGGG	CGCCCTAAACAA	AGGGTG	GTC	ACT	TCT	AGT	CCCAGGA	
17	20	ATGGGAGGG	CGCG	TTCTT	CGTG	GTT	ACT	TTT	AGT	CCCCG
31	25	CLTTGGG	CGTG	AATGTC	CACG	GGT	ACC	TCC	GGT	CCCCAAAGAG
16	30	TCCGGG	CTTCGG	GAT	TCGTG	GTC	ACT	TTC	AGT	CCCGGATATA
32	31									5'TCCGGCGCAAGT
										acttgag3'
18	35	ACGGGAGGG	CACG	TTCTT	CGTG	GTT	ACT	TTT	AGT	CCCCG
5	41	=GGG	CTGAGTa3'	5'LactCAG	GGC	ACTGCAAGCAAT	TGTGGT	CCCCAAT=		
14	47	GTGGGTGGGATCGGG	ATG	CTC	GTC	ACT	TCT	AGT	CCCact	

Table 4. Frequency of base pairs in the helical regions of the consensus motif shown in Figure 1.

5	Position ^a	Base pair ^b						other
		AT	TA	GC	CG	TG	GT	
10	I-1	0	0	21	0	0	0	0
	I-2	0	0	21	0	0	0	0
	I-3	5	0	16	0	0	0	0
	I-4	3	5	1	4	1	0	7
	I-5	2	3	3	4	0	0	9
15	II-1	0	1	2	17	0	0	1
	II-2	5	5	5	1	0	4	1
	II-3	3	4	7	6	0	0	1
	II-4	3	0	8	5	0	0	4
20	III-1	21	0	0	0	0	0	0
	III-2	0	10	0	11	0	0	0
	III-3	0	7	0	13	1	0	0

^aHelices are numbered with roman numerals as shown in Figure 1. Individual base pairs are numbered with Arabic numerals starting with position 1 at the helix junction and increasing with increased distance from the junction.

^b The TG and GT base pairs to the Watson-Crick base pairs for this analysis were included. There is a total of 21 sequences in the set.

Table 5. Affinities of the minimal DNA ligands to PDGF AA, PDGF AB and PDGF BB.

		K_d , nM		
5	<u>Ligand</u>	<u>PDGF AA</u> ^a	<u>PDGF AB</u> ^b	<u>PDGF BB</u> ^b
	20t	47 ± 4	0.147 ± 0.011	0.127 ± 0.031
	36t	72 ± 12	0.094 ± 0.011	0.093 ± 0.009
10	41t	49 ± 8	0.138 ± 0.009	0.129 ± 0.011

^aData points shown in Figure 3A were fitted to eq 1 (Example 1).

15 ^bData points in Figures 3B and 3C were fitted to eq. 2. The dissociation constant (K_d) values shown are for the higher affinity binding component. The mole fraction of DNA that binds to PDGF AB or PDGF BB as the high affinity component ranges between 0.58 to 0.88. The K_d values for the lower affinity interaction range between 13 to 78 nM.

Table 6. Relative affinity for PDGF-AB of ligand 36t variants.

		SEQ		Composition*	$\frac{K_d^{\text{ligand}}}{K_d^{36t}}$ **
Ligand	ID NO				
5	36t	84	CACAGGCTACGGCACGTAGAGCATCACCATGATCCTGTG[3'T]		1.0
	1073	97	<u>CAC</u> AGGCTACGGCACGTAGAGCATCACCATGATCCT <u>GUG</u> [3'T]		11.8
	1074	98	CACAGGCTA <u>CGGCACGU</u> AGAGCATCACCATGATCCTGTG[3'T]		3.1
	1075	99	CACAGGCTACGGCACGTAGAGCATC <u>ACCAU</u> GATCCTGTG[3'T]		10
	1076	100	<u>CAC</u> AGGCTA <u>CGGCACGU</u> AGAGCATC <u>ACCAU</u> GATCCTGTG[3'T]		440
10	1145	101	CACAGGCTACGGCACGTAGAGCAT <u>CACCA</u> TGATCCTGTG[3'T]		0.27
	1148	102	CAC <u>AGGCUA</u> CGGCACGTAGAGCATCACCATGATCCTGTG[3'T]		281
	1144	103	CACAGGCTACGGCACGTAGAGCATCACCAT <u>GAUCCU</u> GTG[3'T]		994
	1142	104	CACAGGCTACGGCACG <u>U</u> AGAGCA <u>AU</u> CACCATGATCCTGTG[3'T]		12.9
	1149	105	CACAGGCTACGGCACGTAG <u>AGC</u> ATCACCATGATCCTGTG[3'T]		2.9
15	EV1	106	CACAGGCTACGGCACGTAG <u>AGC</u> ATCACCATGATCCTGTG[3'T]		35.1
	EV2	107	CACAGGCTACGG <u>CACGU</u> AGAGCATCACCATGATCCTGTG[3'T]		5.3
	EV3	108	CACAGGCTA <u>CGGCAC</u> GTAGAGCATCACCATGATCCTGTG[3'T]		1.5
	EV4	109	<u>CAC</u> AGGCTACGGCACGTAGAGCATCACCATGATCCTGTG[3'T]		4.5
	EV5	110	CACAGGCTACGGCACGTAGAGCATCACCATGATCC <u>UGUG</u> [3'T]		2.3
20	1157	111	CACAGGCTACGGCACGTAGAGCATCACCATGATCCTGTG[3'T]		1.0
	1160	112	CACAGGCTACGGCACGTAGAGCATCACCATGATCCTGTG[3'T]		1.4
	1161	113	CACAGGCTACGGCACGTAGAGCATCACCATGATCCTGTG[3'T]		0.22
	1162	114	CACAGGCTACGGCACGTAGAGCATCACCATGATCCTGTG[3'T]		0.52
	1165	115	CACAGGCTACGGCACGTAGAGCATCACCATGATCCTGTG[3'T]		0.61
25	1164	116	CACAGGCUACGGCACGTAGAGCATCACCATGATCCTGTG[3'T]		0.45
	1166	117	CACAGGCTACGGCACG <u>U</u> AGAGCATCACCATGATCCTGTG[3'T]		0.76
	1159	118	CACAGGCTACGGCACGTAGAGCA <u>U</u> CACCATGATCCTGTG[3'T]		0.37
	1163	119	CACAGGCTACGGCACGTAGAGCATCACCA <u>U</u> GATCCTGTG[3'T]		1.3
	1158	120	CACAGGCTACGGCACGTAGAGCATCACCATGA <u>U</u> CCTGTG[3'T]		2.4
30	1255	121	CACAGGCUACGGCACG <u>U</u> AGAGCA <u>U</u> CACCATGA <u>UCCU</u> GTG[3'T]		24.2
	1257	122	CACAGGCTA <u>CGGCAC</u> GTAGAGCATC <u>ACCA</u> TGATCCT <u>GUG</u> [3'T]		1.3

Table 6 Cont.

		SEQ		
	<u>Ligand</u>	ID NO	<u>Composition*</u>	$K_d^{\text{ligand}}/K_d^{36t} **$
5	1265	123	CACAGGCTA <u>CGGC</u> ACGTAGAGCAT <u>CACCAT</u> TGATCCUGUG[^{3'} T]	1.4
	1266	124	CACAGGCUA <u>CGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUGUG[^{3'} T]	1.0
	1267	125	CACAGGCTA <u>CGGC</u> ACGTAGAGCAT <u>CACCAT</u> TGATCCUGUG[^{3'} T]	4.2
	1269	126	CACAGGCUA <u>CGGC</u> ACGTAGAGCAT <u>CACCAT</u> TGATCCUGUG[^{3'} T]	0.87
	1295	127	CAGGCUA <u>CGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	0.9
10	1296	128	CAGGCU- <u>CGGC</u> ACG-AGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	2.1
	1297	129	CAGGCU- <u>CGGC</u> ACG-AGAGCAUC- <u>CCA</u> -GATCCUG[^{3'} T]	2.9
	1303	130	CAGGCUA <u>CGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	5.8
	1304	131	CAGGCU <u>ACGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	607
	1305	132	CAGGCU <u>ACGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	196
15	1306	133	CAGGCU <u>ACGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	4.4
	1327	134	CAGGCUA <u>CGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	0.63
	1328	135	CAGGCUA <u>CGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	2.2
	1329	136	CAGGCUA <u>CGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	0.72
20	1369	137	CAGGCUA <u>CGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	0.37
	1374	138	CAGGCU <u>ACGGC</u> ACGTAGAGCAUC <u>ACCAT</u> TGATCCUG[^{3'} T]	1.5
	1358	139	CAGGCUA <u>CG</u> -S-CGTAGAGCAUCA-S-TGATCCUG[^{3'} T]	0.54
	1441	140	CAGGCUA <u>CG</u> -S-CGTAGAGCAUCA-S-TGATCCUG[^{3'} T]	0.33

25

*A,C,G,T=deoxy-A,C,G,T; A,C,G,U=2'-OMe-A,C,G,T; C,U=2'-fluoro-C,U;
S=hexaethyleneglycol spacer; [^{3'}T]=inverted (3'-3') T.

**Kd^{36t} value of 0.178±88 pM used for the calculation is average, with standard deviation,

30

of four independent measurements (94±11, 161±24, 155±30 and 302±32 pM).

Table 7. Relative affinity for PDGF-BB of ligand 36ta variants.

The effect of various substitutions on the affinity of ligand 36ta for PDGF-BB.

5	<u>Ligand</u>	<u>Composition*</u>	$\frac{K_d^{\text{ligand}}}{K_d^{36ta}}$ **	SEQ. ID NO.
	36ta	CAGGCTACGGCACGTAGAGCATCACCATGATCCTG[3'T]	1.0	141
	NX21568	CAGGCUAC <u>G</u> -S-CGTAGAGCAUCA-S-TGATCCUG[3'T]	0.63	142
	NX21617	[L1]CAGGCUAC <u>G</u> -S-CGTAGAGCAUCA-S-TGATCCUG[3'T]	0.54	143
10	NX21618	[L1]CAGGCUACG-S-CGTAGAGCAUCA-S-TGAUCCTG[3'T]	418	144
	NX31975	[L2] CAGGCUAC <u>G</u> -S-CGTAGAGCAUCA-S-TGATCCUG[3'T]	0.54	148
	NX31976	[L2] CAGCGUAC <u>G</u> -S-CGTACCGATUCA-S-TGAAGCUG[3'T]		149

*A,C,G,T=deoxy-A,C,G,T; A,G=2'-OMe-A,G; C,U=2'-F-C,U; S=hexaethyleneglycol
 15 spacer (from Spacer Phosphoramidite 18, Glen Research, Sterling, VA); [3'T]=inverted (3'-3') T; [L1]=dT amine (Glen Research, Sterling, VA); [L2] = pentyl amino linker.

** $K_d^{36ta} = 0.159 \pm 13$ pM.

Table 8. Starting RNA and PCR primers for the 2'-fluoropyrimidine RNA SELEX experiment.

		SEQ ID
5		NO
	Starting 2'-fluoropyrimidine RNA:	
	Starting RNA:	
	5'-GGGAGACAAGAAUAACGCUCAA[-50 N-]UUCGACAGGAGGCUCACAACAGGC-3'	36
10	PCR Primer 1:	
	5'-TAATACGACTCACTATAGGGAGACAAGAATAACGCTCAA-3'	37
	PCR Primer 2:	
15	5'-GCCTGTTGTGAGCCTCCTGTCGAA-3'	38

Table 9. Sequences of the evolved region of 2'-fluoropyrimidine RNA high affinity ligands to PDGF AB. Sequences of the fixed region (Table 8) are not shown.

		SEQ ID NO.
5		
	1 CGGUGGCAUUCUUCACUCCUUCUCGCUUUCUCGCGUUGGGCNCGA	39
	2 CCAACCUUCUGUCGGCGUUGCUUUUUGGACGGCACUCAGGCUCCA	40
	3 UCGAUCGGUUGUGUGCCGGACAGCCUUAACCAGGGCUGGGACCGAGGCC	41
10	4 CUGAGUAGGGGAGGAAGUUGAAUCAGUUGUGGCGCCUCUCAUUCGC	42
	5 CAGCACUUUCGCUUUUCAUCAUUUUUUCUUCACUGUUGGGCGCGGAA	43
	6 UCAGUGCUGGCGUCAUGUCUCGAUGGGGAUUUUUCUUCAGCACUUUGCCA	44
	7 UCUACUUUCCAUUUCUCUUUUUCUUCACGAGCGGGUUCAGUGAACCA	45
	8 CGAUAGUGACUACGAUGACGAAGGCCGCGGGUUGGAUGCCCGCAUUGA	46
15	10 GUCGAUACUGGCGACUUGCUCUCCAUUGGCCGAUUAACGAUUCGGUCAG	47
	13 GUGCAAACUUAACCCGGGAACCGCGCGUUCGAUCGACUUCCUUUCCA	48
	15 AUUCCGCGUUCGGAUUAUCCUGUGCUCGGAAAUCGGUAGCCAUAGUGCA	49
	16 CGAACGAGGAGGGAGUGGCAAGGGAUGGUUGGAUAGGCUCUACGCUCA	50
	17 GCGAAACUGGCGACUUGCUCUCCAUUGGCCGAUUAACGAUUCGGUUCAU	51
20	18 CGAACGAGGAGGGAGUCGCAAGGGAUGGUUGGAUAGGCUCUACGCUCAA	52
	19 CGAGAAGUGACUACGAUGACGAAGGCCGCGGGUUGAAUCCCUCAUUGA	53
	20 AAGCAACGAGACCUGACGCCUGAUGUGACUGUGCUUGCACCCGAUUCUG	54
	21 GUGAUUCUCAUUCUCAUUGCUUUCACAAUUCUUUUUCCACUUCAGCGUGA	55
	22 AAGCAACGAGACUCGACGCCUGAUGUGACUGUGCUUGCACCCGAUUCU	56
25	23 UCGAUCGGUUGUGUGCCGGACAGCUUUGACCAUGAGCUGGGACCGAGGCC	57
	24 NGACGNGUGGACCUGACUAAUCGACUGAUCAAAGAUCGCGCCGAGUUGG	58
	26 CACUGCGACUUGCAGAAGCCUUGUGUGGCGGUACCCCUUUGGCCUCG	59
	27 GGUGGCAUUCUUCAUUUUCCUUCUCGCUUUCUCCGCCGUUGGGCGCG	60
	29 CCUGAGUAGGGGGGAAGUUGAAUCAGUUGUGGCGCUCUACUCAUUCGCC	61
30	30 GUCGAAACUGGCGACUUGCUCUCCAUUGGCCGAUUAACGAUUCGGUUCA	62
	31 GCGAUACUGGCGACUUGCUCUCCAUUGGCCGAUUAACGAUUCGGCUCAG	63
	32 ACGUGGGGCACAGGACCGAGAUCCCUCCGGCAUAGCCGCUACCCACC	64
	33 CACAGCCUNANAGGGGGGAAGUUGAAUCAGUUGUGGCGCUCUACUCAUUCGC	65
	34 ANGGGUAUGGUGACUUGCUCUCCAUUGGCCGAUUAACGAUUCGGUCAG	66
35	35 CCUGCGUAGGGNGGGAAGUUGAAUCAGUUGUGGCGCUCUACUCAUUCGCC	67

Table 9 Cont.

			SEQ ID NO.
5	39	CGAACGAGGAGGGAGUGGCAAGGGAUGGUUGGAUAGGCUCUACGCUCA	68
	41	GUGCAAACUUAACCCGGGAACCGCGCGUUUCGAUUCGCUUCCNUAUUCCA	69
	42	CGAACGAGGAGGGAGUGGCAAGGGACGGUNNAUAGGCUCUACGCUCA	70
	43	UCGGUGUGGCUCAGAAACUGACACGCGUGAGCUUCGCACACAUCUGC	71
	44	UAUCGCUUUUCAUCAAUUCCACUUUUUCACUCUNUAACUUGGGCGUGCA	72
10	45	GUGCAAACUUAACCCGGGAACCGCGCGUUUCGAUCCUGCAUCCUUUUUCC	73
	46	UCGNUCGGUUGUGUGCCGGCAGCUUUGUCCAGCGUUGGGCCGAGGCC	74
	47	AGUACCCAUCUCAUCUUUUCCUUUCCUUUCUUAAGGCACAUUGAGGGU	75
	49	CCUGAGUAGGGGGGGAAGUUGAACCAGUUGUGGCNGCCUACUCAUUCNCCA	76
	51	CCNNCCUNCUGUCGGCGCUUGUCUUUUUGGACGGGCAACCCAGGGCUC	77
15	52	CCAACCUNCUGUCGGCGCUUGUCUUUUUGGACGAGCAACUCAAGGCUCGU	78
	53	CCAGCGCAGAUCCCGGGCUGAAGUGACUGCCGGCAACGGCCGCUCCA	79
	54	UUCCCGUAACAACUUUUCAUUUUCACUUUUAUCCAACCAGUGAGCAGCA	80
	55	UAUCGCUUUCAUCAAUUCCACUCCUUCACUUCUUUAACUUGGGCGUGCA	81